

Est.
1841

YORK
ST JOHN
UNIVERSITY

Schwendel, Arved, Death, R. G., Fuller, I. C. and Tonkin, J. D. (2011) A new approach to assess bed stability relevant for invertebrate communities in upland streams. *River Research and Applications*, 28 (10). pp. 1726-1739.

Downloaded from: <https://ray.yorksja.ac.uk/id/eprint/2618/>

The version presented here may differ from the published version or version of record. If you intend to cite from the work you are advised to consult the publisher's version:
<https://doi.org/10.1002/rra.1570>

Research at York St John (RaY) is an institutional repository. It supports the principles of open access by making the research outputs of the University available in digital form. Copyright of the items stored in RaY reside with the authors and/or other copyright owners. Users may access full text items free of charge, and may download a copy for private study or non-commercial research. For further reuse terms, see licence terms governing individual outputs. [Institutional Repositories Policy Statement](#)

RaY

Research at the University of York St John

For more information please contact RaY at
ray@yorksja.ac.uk



A new approach to assess bed stability relevant for invertebrate communities in upland streams

Journal:	<i>River Research and Applications</i>
Manuscript ID:	RRA-10-0147.R1
Wiley - Manuscript type:	Research Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Schwendel, Arved; Massey University, Institute of Natural Resources - Ecology Death, Russell; Massey University, Institute of Natural Resources - Ecology Fuller, Ian; Massey University, Institute of Natural Resources - Geography Tonkin, Jonathan; Massey University, Institute of Natural Resources - Ecology
Keywords:	bedload transport, lotic ecosystem, mountain stream, stream ecology, substrate stability, tracer stone

SCHOLARONE™
Manuscripts

1
2
3 **A new approach to assess bed stability relevant for invertebrate communities in**
4 **upland streams**
5
6

7
8
9 Arved C. Schwendel, Ecology Group, Institute of Natural Resources, Massey
10 University, Private Bag 11222, Palmerston North 4442, New Zealand
11

12
13
14 *Current address:* College of Life and Environmental Sciences - Geography,
15 University of Exeter, Exeter EX4 4RJ, UK
16

17 Email: A.Schwendel@exeter.ac.uk

18
19 Phone: +44 1392724488
20

21 Fax: +44 1392723342
22
23

24
25 Russell G. Death, Ecology Group, Institute of Natural Resources, Massey University,
26 Palmerston North, New Zealand, Email: R.G.Death@massey.ac.nz
27

28
29
30 Ian C. Fuller, Physical Geography Group, Institute of Natural Resources, Massey
31 University, Palmerston North, Email: I.C.Fuller@massey.ac.nz
32

33
34
35 Jonathan Tonkin, Ecology Group, Institute of Natural Resources, Massey University,
36 Palmerston North, New Zealand, Email: J.Tonkin@massey.ac.nz
37
38
39
40
41
42
43

44 **Running head:** A new approach to assess bed stability
45
46
47
48

49 **Keywords:** bedload transport, lotic ecosystem, mountain stream, stream ecology,
50 substrate stability, tracer stone
51
52
53
54
55
56
57
58
59
60

ABSTRACT

1
2
3
4
5
6
7 Composition and structure of lotic ecosystems can be affected by substrate instability.
8
9 Consequently stream ecologists have used various methods to determine bed stability
10 characteristics. However, the link between community composition and these
11 measurements varies because benthic biota often respond to combinations of bed
12 stability characteristics. This paper presents a protocol to determine reach-scale
13 stream bed stability in mountain streams which is relevant for invertebrate
14 communities (Stream Bed Stability for Invertebrates, SBSI). The approach is
15 calibrated on community composition response to bed stability but does not measure
16 any single bed stability characteristic *per se*. It consists of 13 parameters that are
17 assessed once at each reach with minimal instrumentation and low interference with
18 the substrate. These 13 parameters cover aspects of sediment supply from banks,
19 transport capacity and substrate erodibility as well as effects of particle transport on
20 channel bottom structures, substrate assemblage and single grains. Application of the
21 SBSI protocol improved the relationship between bed stability and community
22 diversity compared to when conventional bed stability measures were employed. The
23 SBSI protocol provides a cost- and time-effective assessment method for bed stability
24 and its application can facilitate research on invertebrate community response to
25 physical disturbance.
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

INTRODUCTION

Flow influences many important structural attributes of stream ecosystems such as substrate stability, habitat volume and channel morphology (Poff and Ward, 1989). Variation in discharge is recognised as one of the fundamental determinants of structure and function of benthic communities in lotic ecosystems (Resh *et al.*, 1988; Reice *et al.*, 1990; Lake, 2000; Death, 2008). Floods can cause movement of coarse bed substrate which can affect composition of periphyton (Biggs *et al.*, 1999), invertebrate (Cobb *et al.*, 1992; Death and Winterbourn, 1995; Holomuzki and Biggs, 2000), bryophyte (Suren and Duncan, 1999) and macrophyte communities (Riis *et al.*, 2008). However, different groups of biota respond to different aspects of bed stability on a range of scales. For instance the reaction to patchy scour or fill varied between invertebrate taxa while on a larger scale stable patches might mitigate the effects substrate instability (Matthaei and Townsend, 2000). Bed stability is a characteristic feature of alluvial channels comprising aspects like entrainment, transport and deposition of substrate as well as abrasion by suspended material on scales ranging from a single particle to an entire reach. These bed stability characteristics might affect sessile organisms in different ways than more mobile groups of biota (Downes, 1990; Englund, 1991; Holomuzki and Biggs, 2000; McAuliffe, 1984). Consequently some methods to quantify bed stability perform well with one group of organisms but show only a weak connection with other groups (Duncan *et al.*, 1999; Schwendel *et al.*, 2011a). This in turn is reflected in the wide variety of bed stability measurements used by stream ecologists to examine the effects of flow disturbance (Schwendel *et al.*, 2010).

The effects of substrate movement on stream invertebrate communities via habitat alteration, displacement and death of individuals, and changes in their food sources are widely recognised (e.g. Townsend *et al.*, 1997; Matthaei and Townsend, 2000; Effenberger *et al.*, 2006; Death, 2008; Schwendel *et al.*, 2011b). Different levels of bed stability, e.g. apparent in depth and pattern of disturbance or in transport distance of particles, are reflected in invertebrate community composition for instance via recolonisation abilities of individual taxa (Death, 2008). The methods employed to assess bed stability in relation to invertebrate community metrics are reviewed in Schwendel *et al.* (2010) and include calculation of critical shear stress (Newbury, 1984; Cobb *et al.*, 1992; Death and Winterbourn, 1995), FST-hemispheres (Dittrich

1
2
3 and Schmedtje, 1995; Merigoux and Doledec, 2004), scour chains (Palmer *et al.*,
4 1992; Matthaei and Townsend, 2000; Effenberger *et al.*, 2006), scour plates (Palmer
5 *et al.*, 1992), tracer stones (Death and Winterbourn, 1994; Townsend *et al.*, 1997;
6 Death and Zimmermann, 2005; Barquin and Death, 2006), morphological budgeting
7 (Schwendel *et al.*, 2011a) and the Pfankuch Stability Index (Pfankuch, 1975; Death
8 and Winterbourn, 1995; Townsend *et al.*, 1997; Death, 2002). Each of these methods
9 can only assess a distinct set of bed stability characteristics and the strength of the
10 relationship with invertebrate diversity and community composition varies
11 (Schwendel *et al.*, 2011a). The need of site specific calibration (e.g. bedload transport
12 formulae and acoustics sensors) and interference with the substrate (e.g. scour plates
13 and bedload traps) can constrain application for multi site studies and concomitant
14 invertebrate sampling respectively (Schwendel *et al.*, 2010). Insufficient spatial (e.g.
15 bedload samplers) or temporal coverage (e.g. FST-hemispheres) for reach-wide, long-
16 term bed stability assessment can be an additional problem. Further, time and cost
17 constraints can often prevent application of elaborate methods. Visual surveys of
18 stream bed properties such as the Pfankuch Stability Index can circumvent some of
19 these limitations but they can potentially be biased by observers or regional factors
20 such as substrate lithology.

21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
Thus a technique that combines the strengths of elaborate bed stability
measurements with the easy application of a visual approach would facilitate research
on stream invertebrates and increase comparability between studies. Consequently,
this paper presents a straightforward survey protocol specifically calibrated for the
assessment of reach-scale stream bed stability relevant for invertebrate community
composition (SBSI). It needs to be pointed out that the SBSI survey does not measure
any single aspect of bed stability *per se* but determines a characteristic response of
invertebrate community composition to a combination of bed stability characteristics.
The SBSI was validated at independent sites using *in situ* marked tracer stones and the
bottom component of the Pfankuch Index, two techniques that were shown to be well
related to invertebrate community metrics (Schwendel *et al.*, 2011a). Additionally the
connection between bed stability measured with SBSI and community metrics was
explored.

Application for the SBSI method may include scientific studies of disturbance-
diversity relationships and habitat characteristics as well as assessment of the
potentially confounding effects of bed instability on invertebrate community

1
2
3 composition when the latter is employed to determine water quality or environmental
4 status of a stream.
5
6
7

8 9 METHODS

10 11 *Study sites*

12
13
14
15
16 Data for calibration and validation of SBSI protocol were collected between October
17 2007 and March 2010 from 54 mountain stream reaches in the southern part of the
18 North Island of New Zealand. They were located in the axial Tararua (n = 12) and
19 Ruahine Ranges (n = 11), the Central Volcanic Plateau (n = 13) and around Mt.
20 Egmont (n = 18) (Figure 1). The former ranges consist of uplifted folded and faulted
21 Mesozoic greywacke and argillite whereas the other mountains are composed of
22 Quaternary andesitic volcanic deposits. Catchment vegetation was dominated by
23 native broadleaf-podocarp forests, scrub and tussock grassland and anthropogenic
24 influence is relatively small (e.g. <0.1% urban land use, 0-45% non-intensive pasture
25 and no infrastructure upstream of sites). Consequently water quality was expected to
26 be relatively unimpaired. The studied stream reaches varied considerably in substrate
27 assemblage, width, channel form (Table I) and sediment supply (Schwendel and
28 Fuller, in press). Substrate composition ranged between gravels and cobbles although
29 some sites contained a considerable proportion of boulders. Riparian vegetation was
30 variable with native forest, willows and poplars, native scrub, non-intensive pasture,
31 tussock and bare ground present. Some of the reaches were laterally confined by
32 vegetated banks, whereas others migrated within a wide active channel zone.
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47

48 49 *Invertebrate communities*

50
51 Five Surber samples (500 μm mesh, 0.1 m^2) were collected from riffles during periods
52 of baseflow at least two weeks after the last spate to ensure a characteristic species
53 assemblage was collected. Seasonal variability in New Zealand stream invertebrate
54 communities is generally low (Towns, 1981; Winterbourn, 2000) however, this was
55 tested and confirmed at 18 of the sites where samples were taken three times
56 throughout the year (Schwendel *et al.*, 2011a and J. Tonkin, unpublished data).
57
58 Samples were stored in 4% formalin or >60% isopropyl alcohol and later sorted. Very
59
60

1
2
3 abundant taxa (>300 individuals per sample) were subsampled following Vinson and
4 Hawkins (1996): samples expected to contain large numbers were divided in equal
5 subsamples of which one was initially searched for invertebrates. Only those taxa
6 which number of individuals did not exceed 300 in the first subsample were searched
7 for in the second subsample. Invertebrates were identified to the lowest possible
8 taxonomic level using the keys in McFarlane (1951), Winterbourn (1973), Towns and
9 Peters (1996) and Winterbourn *et al.* (2006). Invertebrates were sampled where
10 applicable from riffles because community composition there is likely to reflect
11 gradations in substrate stability and on a larger scale instability in riffles affects also
12 pools, e.g. via bedload transport.
13
14
15
16
17
18
19
20
21

22 *Periphyton and habitat parameters*

23
24
25
26 At each invertebrate sampling point depth, wetted stream width and near-bottom flow
27 velocity were measured. The latter was recorded over 60 s with an electromagnetic
28 flow meter (Model 801, Valeport Ltd., Devon, UK) 0.05 m above the stream bed. At
29 each site pH and temperature corrected conductivity were measured using Eutech
30 pHtestr2 and ECScan Low+ (Eutech Instruments, Singapore) respectively. Percentage
31 aerial cover of riparian land use categories (native vegetation, pasture and willows)
32 within a strip of approximately 5 m and the fraction of dry active channel bare of
33 vegetation under base flow conditions was estimated visually.
34
35
36
37
38
39

40
41 Chlorophyll a pigment concentration on five gravel-sized stones that were
42 collected beside invertebrate samples was assessed as a measure of periphyton
43 biomass. The stones were transported in the dark in cooled stream water before
44 storing them at -18° C. Pigments were extracted in 90% acetone for 18 h at 5° C in the
45 dark before the chlorophyll a absorption was measured using a Cary 50 Conc UV-
46 Visible spectrometer (Varian, Mulgrave, Australia). Chlorophyll a pigment
47 concentration was calculated (Steinman and Lamberti, 1996; APHA, 1998) and
48 corrected for stone surface area which was estimated based on measurement of the a-,
49 b- and c-axes of the gravels with a sliding calliper following Graham *et al.* (1988).
50
51
52
53
54
55

56
57 Substrate composition of riffles was assessed by measuring the b-axis of >100
58 randomly collected particles (Wolman, 1954) and classifying them according to a
59 modified Wentworth scale.
60

Bed stability

Substrate stability was assessed with two established reference measures: tracer particles and the Pfankuch Stability Index. For the development of the new approach a set of 38 candidate variables (Table II) were selected from a large array of parameters potentially related to stream bed stability (Knighton, 2008; Petts and Foster, 1985) in respect to importance and practicability of assessment with minimal instrumentation in the field. These candidate variables were evaluated at stream sections with a length of approximately 5 – 7 times stream width to include, where present at least one riffle-pool unit (Keller and Melhorn, 1978).

Candidate variables are associated with the riparian environment (denoted A), the cross (B) and longitudinal profile (C) of the channel, the channel bottom structure (D) and the substrate (E). The density and composition of the riparian vegetation within a 5 m-strip along the active channel zone (A1, A3) reflects bank stabilisation by roots, pressure from land use and frequency and magnitude of flood disturbance. Together with bank erosion (B2) and deposition of derived fine sediments (B3) these parameters indicate sediment supply from banks and slopes. These processes influence substrate characteristics (E3-6) which can be relevant for bed stability. Transport capacity is assessed in terms of available potential energy (slope) (C1), expenditure on roughness elements (D6), channel adjustments (C2, D4) and flood regime (A2). The channel dynamics resulting from sediment supply and transport capacity are reflected in channel form (B1), structures (D1, D3-5), aquatic vegetation (D2) and substrate characteristics (E1-4, E7). Additionally lithology of the substrate, weather (sunny, overcast or rain) and state of the floodplain substrate (dry or wet) were recorded because these factors could potentially interfere with visual evaluation methods such as the Pfankuch Index (A. C. Schwendel, unpublished data).

Tracer particles were used to assess stream bed stability. Five randomly selected tracer stones in each of three size classes (D_{50} , D_{70} and D_{90}) were marked with radio-frequency identification (RFID) tags (23 mm glass tags, Texas Instruments, Dallas, USA) which were attached *in situ* to stones in riffles using wet curing epoxy-concrete (K273, Nuplex Construction Products, Hamilton, New Zealand). When high turbulence or fast flow velocity prevented underwater application (11% of particles), stones were removed from the river bed for tag attachment and afterwards carefully re-embedded. The percentage of entrained *in situ*-marked tracer stones and re-

1
2
3 embedded tracers was significantly correlated (Spearman-rank correlation, $R = 0.70$,
4 $df = 26$, $p = 0.0001$). Relocation and identification of each tracer stone was carried out
5 contactless using a portable antenna and datalogger (OregonRFID, Portland, USA).
6 Initial and subsequent positions of tagged stones were surveyed using high precision
7 differential GPS or marked on riparian vegetation and stable banks. Relocation
8 surveys took place approximately every two months or after high discharge events
9 over a total period of six months. The entire bed and active channel downstream of
10 the last position of each tracer particle was searched intensively to the next local
11 sediment trap (e.g. riffle) beyond a minimal distance of 50 m. Stones that could not be
12 recovered were assigned a travel distance of 50 m. Although this was less than usually
13 searched it accounted for tracers lost by deep burial (>0.6 m), storage in inactive parts
14 of the floodplain, tag damage and malfunction. The travelled distance of the tracer
15 particles was converted to an index of bed stability (TTM = sum of tracer movement)
16 using the following approach:
17
18
19
20
21
22
23
24
25
26
27
28
29

$$30 \quad TTM = (d_{50} * s_{50} / n_{50} + d_{70} * s_{70} / n_{70} + d_{90} * s_{90} / n_{90}) / (d_{50} + d_{70} + d_{90}) \quad (1).$$

31
32

33 The sum of the moved distance s of stones of a size class between the surveys is
34 divided by the counted recoveries n and weighted by the geometric mean particle size
35 d of that class.
36
37

38 As a second independent measure of bed stability the bottom component of the
39 Pfankuch Stability Index (BCP) (Pfankuch, 1975) was employed once at each site.
40 The bottom component was preferred over the total index because in previous studies
41 it showed a better relationship with other measures of bed stability (Death and
42 Winterbourn, 1994) and is well related to biological data (Death and Winterbourn,
43 1995; Suren, 1996). It involves allocation of an observer's subjective visual
44 evaluation of six attributes, including substrate brightness, angularity, consolidation of
45 particles, percentage of stable materials, evidence of scouring and state of clinging
46 aquatic vegetation, to four predetermined categories to which scores are weighted
47 according to their perceived importance. The sum of the scores results in a stability
48 index, where high values represent low stability.
49
50
51
52
53
54
55
56
57
58
59
60

Data analysis

1
2
3 The collected data were examined in four steps: (1) analysis of invertebrate
4 community composition and structure, (2) development of the SBSI protocol, (3)
5 exploration of the relationship between SBSI, other measures of bed stability and
6 community metrics and (4) validation of the SBSI protocol at independent sites in
7 respect to other bed stability measures and relevance for invertebrate communities.
8
9

10
11
12 The composition of the invertebrate community at 46 calibration sites (Figure 1)
13 was explored with non-metric multidimensional scaling (NMDS) in PC-ORD 5.0
14 (MjM Software, Gleneden Beach, USA) using standardised (by maximum)
15 invertebrate taxa abundance. Association of the derived axis scores with measured
16 environmental parameters and selected variables from the Freshwater Environments
17 of New Zealand (FWENZ) database (Wild *et al.*, 2005) was assessed using Pearson's
18 correlation. The axis that was best correlated to conventional bed stability measures
19 TTM and BCP was selected for calibration of the SBSI. Community diversity
20 (Brillouin Index), taxa number, rarefied taxa number (for 200 individuals following
21 Sanders (1968) and Hurlbert (1971)) and mean number of individuals per 0.1 m² were
22 calculated for all sites in PRIMER v6 (Plymouth Marine Laboratory, Plymouth, UK).
23
24
25
26
27
28
29
30
31

32 The SBSI was developed with linear best subset regression (Statistix 9.0,
33 Analytical Software, Tallahassee, USA) using the selected NMDS axis as dependent
34 variable and the 38 parameters assessed in the field (Table II) as independent
35 variables. Adjusted R², residual mean square error, Mallows' Cp, predicted residual
36 sum of squares and Akaike's Information Criterion for small samples (AICc) were
37 used to compare models.
38
39
40
41

42 The relationship between the SBSI site scores, bed stability measured with
43 tracer stones and the bottom component of the Pfankuch Index, and invertebrate
44 community metrics was assessed with Spearman rank correlation to account for the
45 non-normal distribution of variables. This was accomplished for the 46 sites used for
46 SBSI calibration to show the relevance of the SBSI for invertebrate communities and
47 separately for the eight validation sites. The latter consisted of four randomly selected
48 reaches in each of the two bedrock regions (volcanic and sedimentary) in order to
49 account for variations in shape and colour of the substrate. Significance from the
50 multiple correlations was adjusted using false discovery rate correction (Benjamini
51 and Hochberg, 1995).
52
53
54
55
56
57
58
59
60

RESULTS

Invertebrate community

A total of 127 invertebrate taxa were collected across the 46 SBSI calibration sites with a mean number of individuals per 0.1 m² of 194 consisting of on average 33 taxa. Overall Trichoptera comprised the largest number of taxa (35%), followed by Diptera (25%) but the samples were numerically dominated by Ephemeroptera larvae (45% of individuals) of which *Deleatidium* was most common (100% of sites) and abundant (42% of individuals).

Ordination (2D stress 0.16) revealed that only one axis was strongly correlated with bed stability measured with tracer stones and the bottom component of the Pfankuch Index (Table III). This axis was also associated with periphyton biomass and the fraction of the active channel bare of vegetation (Figure 2). It was subsequently used to calibrate the SBSI. Sites associated with low bed stability were found in the Ruahine Ranges and around Mt. Egmont and were dominated by *Deleatidium*. In contrast very stable sites were located mostly on the Central Plateau and had a richer fauna and higher number of individuals.

SBSI protocol

Any intercorrelated variables of assessed reach properties were removed from further analysis (Table II). Weather and substrate surface wetness were not significantly correlated with other variables but substrate lithology (andesite and greywacke) was significantly correlated to grain angularity (E11) (Spearman's R 0.82, df = 45, p = 0.0001). Andesitic stones were more rounded than greywacke clasts prior to fluvial transport. Consequently scores for grain angularity were raised by one class at sites with greywacke dominated substrate. Best subset regression, using the NMDS axis best correlated to bed stability measures as dependent variable and the refined set of reach properties as independent variables, led to the identification of an optimal model (Table IV). This model of stream bed stability relevant for invertebrates (SBSI) comprises 13 variables which reflect mostly direct effects of channel dynamics observed on the banks and at the channel bottom. Sediment supply and transport capacity are represented with two variables each which are assessed on the banks and the longitudinal channel profile. Substrate parameters (size and compaction)

1
2
3 constitute a second group mirroring effects of sediment dynamics such as sorting.
4
5 Low variance inflation factors (VIF) indicated that collinearity between the variables
6
7 is low.

8
9 Based on the regression model a field sheet (Appendix 1) was designed that
10 facilitates recording of the variables and allows with the help of a pocket calculator
11 rapid on-site assessment of bed stability. Channel, bank and substrate properties are to
12 be recorded, noted in relevant fields and multiplied with their respective coefficient.
13
14 The sum for each compartment (e.g. banks, longitudinal profile, channel bottom and
15 substrate) is recorded on the right hand side of the sheet and this column is then added
16
17 up to result in the SBSI site score.
18
19
20
21

22 *Bed stability and community metrics*

23
24
25
26 Correlation between the SBSI site scores and community diversity (Brillouin Index),
27
28 taxa number, rarefied taxa number and mean number of individuals was highly
29
30 significant (Table V). These community metrics were also correlated with bed
31
32 stability measured with tracers (except taxa number) or the bottom component of the
33
34 Pfankuch Index but the connection was always weaker than with the SBSI.

35
36 The three measures of bed stability were intercorrelated with the strongest
37
38 relationship apparent between the bottom component of the Pfankuch Index and SBSI
39
40 site scores (Table VI).

41 *Validation at independent sites*

42
43
44
45
46 At eight randomly selected sites a linear relationship was found between bed stability
47
48 assessed with the bottom component of the Pfankuch Index and the SBSI protocol
49
50 (Table VI). In contrast the tracer measure was not correlated with any of the two
51
52 former, however, correlation coefficients were similar or higher than at the sites used
53
54 for SBSI calibration and the failure of detection of a significant relationship might be
55
56 due to the low number of sites. Correlation between the Brillouin Index and SBSI site
57
58 scores was stronger than with any of the other bed stability measures (Table V). In
59
60 contrast taxa number, rarefied taxa number and the mean number of individuals were
slightly better related to the bottom component of the Pfankuch Index.

DISCUSSION

The presented protocol for assessment of bed stability relevant for invertebrates (SBSI) produces site scores highly related to invertebrate community diversity and structure. This connection is stronger than that of any traditional bed stability measure with community metrics at the calibration sites. The SBSI method is calibrated on the response of invertebrate communities, signified by a NMDS axis, to varying degrees of bed stability as measured with traditional techniques and compares well to the NMDS calibration axis (Table VI, Figure 3). The NMDS axis used for calibration of the SBSI is strongly associated with bed stability measures and periphyton biomass. Periphyton as a potential food source for invertebrates influences invertebrate community composition (Death, 2002) but biomass itself is affected by bed movement and can consequently be seen as a proxy for bed stability. The link of the NMDS calibration axis with the percentage of bare active channel reflects the flood regime which influences bed stability. Lack of vegetation on the banks can indicate regular inundation with flows competent to strip vegetation and to prevent perennial plant growth. Alternatively it can be caused by active bank erosion during lower discharges when undercutting of banks can lead to failure. This reflects a high degree of channel activity and sediment input and accordingly bed disturbance. Hence it is reasonable to interpret the NMDS axis as being dominated by bed stability.

Validation at independent sites showed the applicability of the SBSI approach and its relevance for invertebrates. Connection with community diversity is improved when the SBSI is used compared to other bed stability measures but the bottom component of the Pfankuch Index performs slightly better with number of taxa and individuals (Figure 4). However, the SBSI approach can account for regional variation in parameters such as lithology and should be less affected by observer subjectivity than the purely visual assessment of the Pfankuch Index.

The parameters of the SBSI model are summarised in Table VII. Theoretically the total SBSI score ranges between 19 (stable) and 201 (unstable) when extreme values for all parameters are assumed. However, the calibration sites which, according to the bed stability measurements, include both very stable and unstable reaches, cover a range of only 62 to 88. Thus values higher than 80 represent sites with low bed stability whereas SBSI smaller than 70 indicates high bed stability. The substrate sand fraction and homogeneity are potentially the most powerful parameters but their

1
2
3 extreme values seldom occur in mountain streams. At the calibration sites bank
4 vegetation cover and abundance of multiple barforms had the highest mean scores
5 (10.8 and 9.1 respectively) while slope, area of multiple barforms and sand fraction
6 achieved lowest mean scores (<2.3). In the following section for each parameter the
7 relation to bed stability is explored and assessment in the field with the help of the
8 provided field sheet (Appendix 1) is described.
9

10
11
12 Friction slope determines the total energy available for transport and
13 entrainment of particles in a stream. Water surface or stream bed gradient is often
14 used as a surrogate because it is easier to measure (Schwendel *et al.*, 2010). When
15 the ratio of flow depth to roughness element height is high (e.g. during high
16 discharge) this is an acceptable first-order approximation. Bed slope can be estimated
17 in the field, if necessary with the help of an Abney level.
18

19
20 The active channel includes the zone that is dry at baseflow stage but is subject
21 to regular inundation. It is well coupled to the channel and it is involved in processes
22 of sediment transport. In the field this zone can be determined by the absence or
23 scarcity of perennial vegetation and the presence of recent flood debris. The ratio of
24 the active channel width to wetted baseflow channel width is low (e.g. close to 1) for
25 hydrologically stable streams with small variation in flows (e.g. lake fed). With
26 increasing frequency and magnitude of floods a higher ratio is expected although local
27 geomorphology can interfere (e.g. narrow valleys, bedrock constrictions and bank
28 composition). Both this parameter and stream bed slope quantify potential transport
29 capacity and are expressed on a continuous scale. Considering the potential range of
30 values, bed slope has much less weight than the active channel to baseflow channel
31 width ratio in the regression model.
32

33
34 The sediment supply from banks and lateral channel erosion is represented by
35 the categorical parameter bank erosion. It is evaluated in the field on a scale ranging
36 from none over weak and moderate to strong. Strong bank erosion means that eroded
37 surfaces or collapsed banks are present throughout the reach and that lateral erosion is
38 severe. Moderate bank erosion depicts a state where either light and discontinuous
39 bank erosion is common or locally bank erosion is strong. The category “weak bank
40 erosion” is chosen when only patchy and light bank erosion occurs. Extrinsic causes
41 for bank collapse such as trampling cattle or human interference are included in this
42 parameter and are not separately assessed.
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

The percentage of riparian vegetation cover of the upper banks (above bankfull stage) specifies average vegetation density of the understory (e.g. stems per m²), not the canopy cover along both sides of the reach. It was expected to be positively related to bed stability because vegetation reduces surface erosion and dense roots stabilise the banks. However, regression showed an inverse relationship to bed stability which can be explained by land use, altitude aspects and bank composition. The sites with low bank vegetation cover were either in high altitude locations on the Central Volcanic Plateau or natural vegetation was scarce. Anthropogenic land use practices like forestry or gravel mining on floodplains can cause low density of bank vegetation. They are only profitable on relatively stable ground thus reflecting bank stability. Altitude mirrors catchment size and is thus related to stream power. Hence high altitude sites above the tree line with low vegetation cover have usually more stable upper banks than low altitude sites. This parameter combines these two causes of bank vegetation cover while bank protection by roots is obviously of less importance on the infrequent flood-affected upper banks. We used an accuracy of 5% for bank cover estimations.

Substrate size distribution reflects erosion, sedimentation and transport processes. Fine particles require less shear force for selective entrainment than coarse grains. Hiding and protrusion effects can prevent selective entrainment but visual surface substrate assemblage assessment does usually capture only patches dominated by sand and not hiding sand grains between larger particles. Thus the percentage of sand and smaller grain sizes present and the associated low critical shear stress can indicate high sediment mobility given sufficient transport capacity. Erosion and sedimentation of sandy substrate and associated changes in habitat can cause shifts in invertebrate community composition (Palmer *et al.*, 1992; Downes *et al.*, 2006).

Substrate size homogeneity can be caused by sorting (e.g. downstream fining) but depends also on catchment substrate lithology and sediment sources (reworking of older alluvial deposits, hillslope collapses or fresh tributary inputs). However, in mountain streams where substrate variety is usually limited by catchment size, sorting can be instrumental for substrate size composition. Because sorting processes require substrate movement the parameter “substrate homogeneity” is positively related to instability. In the field it requires estimation of the percentage cover of the size classes silt (<0.063 mm), sand (0.063 mm – 2 mm), gravel (2 mm – 64 mm), cobble (64 mm

1
2
3 – 256 mm) and boulder (>256 mm). Then the aerial cover fraction of the dominant
4 size class is divided by the number of classes present.
5

6
7 Packing and compaction of particles is highly developed in stable substrate
8 channels. It can be an effect of incompetent flows or lack of sediment supply. This
9 parameter should not be confounded with overlap of particles because of the stone
10 shape of some lithologies. It can easily be tested by walking in the bed and four
11 categories are distinguished. Tight packing means that in the entire channel stones
12 move only minimally when full body weight is applied and includes bedrock. Wedged
13 packing depicts conditions where only parts of the channel have tight packing or
14 where the entire substrate moves under the foot but does not principally change
15 position (e.g. is entrained afterwards). The “moderately loose” category includes a
16 mix of all four categories throughout the channel skewed towards looser conditions.
17 Stones may change position when stepped on but should not be entirely dislodged.
18 Loose packing means that the foot sinks into the substrate and particles move easily.
19
20
21
22
23
24
25
26
27

28 The categorical parameter “Constitution of particle surface” has been modified
29 from the categories of brightness defined by Pfankuch (1975). It incorporates surface
30 roughness and brightness which can be effects of particle movement. However, it
31 needs to be distinguished between different lithologies (e.g. limestone and volcanic
32 rocks) which have varying spectra of colours and brightness. Particles of different
33 geological origin can have variable surface roughness after the same transport length.
34 Stains and plant growth on stones are dependent on temperature, light, nutrient levels
35 and mineralisation. It is also advisable to allow for weather conditions and surface
36 moisture when stones on the floodplain are investigated: Wet surfaces on a rainy day
37 can appear much duller than in dry and sunny conditions. The categories range from
38 more than 95% of stained particles with considerable organic film and growth, over
39 “65 – 95% dull” and “35 – 65% dull” to less than 35% dull.
40
41
42
43
44
45
46
47
48
49

50 The parameter “Grain angularity” was also adopted from the Pfankuch Index. It
51 ideally expresses the amount of work performed on a particle during fluvial transport
52 but the characteristic depends very much on lithology in terms of hardness,
53 cleavability, stratification and mineral content as well as distance from source. Thus
54 adjustment of the scores of sharp and angular rock types such as mudstone greywacke
55 to the scores of particles that are already rounded prior to fluvial transport (e.g. some
56 volcanic rocks) by the observer is recommended. The categories include particles well
57 rounded in all dimensions with smooth surfaces, corners and edges well rounded in
58
59
60

1
2
3 two dimensions, corners and edges rounded combined with flat surfaces and sharp
4 edges and corners with roughened surfaces.
5
6

7 The percentage of reworked area describes the amount of obvious recent erosion
8 (e.g. bright sections) and sedimentation (bars of fines, filled pools) of the channel
9 bottom. A fraction of more than 80% is rated as very high, 50 – 80% as high, 20 –
10 49% as intermediate and less than 20% as low.
11
12

13 Multiple barforms are a feature of dynamic channels able to adjust to changing
14 sediment supply and floods. However, over short-term they can be relatively stable
15 channel structures creating various habitats and providing potential refugia during
16 smaller spates. Surprisingly, the number of multiple barforms is positively related to
17 bed stability in the SBSI model which might reflect habitat heterogeneity. In contrast
18 their size as a fraction of the total bed area decreases with SBSI bed stability because
19 large areas of multiple barforms indicate substantial channel dynamics. The number
20 of multiple barforms is classified in six categories which are indexed from 0 to 5.
21
22

23 Bedform clusters locally influence flow turbulence causing expenditure of
24 energy which is not available to entrain substrate. They are commonly thought to be
25 resistant to entrainment during high-discharge events (de Jong, 1992; Reid *et al.*,
26 1992) but depending on flood magnitude bed form clusters can be as unstable as
27 single surface stones (Matthaei and Huber, 2002). Thus their suitability as refugia for
28 invertebrates and periphyton varies and they do not necessarily support richer
29 invertebrate faunas because of increased habitat heterogeneity (Biggs *et al.*, 1997;
30 Francoeur *et al.*, 1998; Matthaei and Huber, 2002). For the SBSI protocol abundance
31 of bedform clusters is estimated in the field visually and categorised in four classes
32 ranging from none to abundant (e.g. > 5% aerial cover).
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47

48 CONCLUSIONS

49
50
51 The method presented here for reach-scale assessment of bed stability relevant for
52 invertebrate communities in upland streams seeks to combine statistically derived
53 relationships between bed stability characteristics and the invertebrate community and
54 causal connections. This distinguishes it from other approaches which aim to measure
55 characteristics of bed stability *per se* but often are not very well related to responses
56 of different groups of biota. The SBSI protocol provides a similar or stronger
57 connection with community diversity and composition than traditional bed stability
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

measures. Index calibration was conducted in upland streams to avoid the confounding effects of water quality on invertebrate communities but potentially the SBSI protocol could be applied to a wide range of streams. The SBSI method is straightforward, cost- and time-effective and requires minimal instrumentation (Abney level and pocket calculator) and only one site visit is necessary. Interference with the substrate is low which facilitates concomitant invertebrate sampling and the stability score can be calculated on-site. It should suffer less from difficulties of purely visual assessments (such as the Pfankuch Index) and can account for regional differences (e.g. in lithology). However, observer bias potentially can be a problem. This and applicability at independent sites need to be tested to allow analysis of deficits and adjustments.

ACKNOWLEDGEMENTS

We thank Ernslaw One Limited and other land owners for site access. We want to acknowledge two anonymous reviewers for their helpful comments on the manuscript.

REFERENCES

- APHA. 1998. *Standard methods for the examination of water and wastewater. 20th ed.* American Public Health Association. American Water Works Association, Water Pollution Control Federation.: Washington D.C.
- Barquin J, Death RG. 2006. Spatial patterns of macroinvertebrate diversity in New Zealand springbrooks and rhithral streams. *Journal of the North American Benthological Society* **25**: 768-786.
- Benjamini Y, Hochberg Y. 1995. Controlling the False Discovery Rate. *Journal of Royal Statistical Society. Series B (Methodological)* **57**: 289-300.
- Biggs BJE, Duncan MJ, Francoeur SN, Meyer WD. 1997. Physical characterisation of microform bed cluster refugia in 12 headwater streams, New Zealand. *New Zealand Journal of Marine and Freshwater Research* **31**: 413-422.
- Biggs BJF, Smith RA, Duncan MJ. 1999. Velocity and sediment disturbance of periphyton in headwater streams: biomass and metabolism. *Journal of the North American Benthological Society* **18**: 222-241.

- 1
2
3 Cobb DG, Galloway TD, Flannagan JF. 1992. Effects of discharge and substrate
4 stability on density and species composition of stream insects. *Canadian*
5 *Journal of Fisheries and Aquatic Sciences* **49**: 1788-1795.
6
7
8
9 Death RG. 2002. Predicting invertebrate diversity from disturbance regimes in forest
10 streams. *Oikos* **97**: 18-30.
11
12 Death RG. 2008. Effects of floods on aquatic invertebrate communities. In: Lancaster
13 J, Briers RA (Editors). *Aquatic Insects: Challenges to Populations*. CAB
14 International, Wallingford, Oxfordshire, pp. 103 - 121.
15
16
17 Death RG, Winterbourn MJ. 1994. Environmental stability and community
18 persistence - a multivariate perspective. *Journal of the North American*
19 *Benthological Society* **13**: 125-139.
20
21
22 Death RG, Winterbourn MJ. 1995. Diversity patterns in stream benthic invertebrate
23 communities - the influence of habitat stability. *Ecology* **76**: 1446-1460.
24
25
26 Death RG, Zimmermann EM. 2005. Interaction between disturbance and primary
27 productivity in determining stream invertebrate diversity. *Oikos* **111**: 392-402.
28
29
30 de Jong C. 1992. A reappraisal of the significance of obstacle clasts in cluster
31 bedform dispersal. *Earth Surface Processes and Landforms* **16**: 737-744.
32
33
34 Dittrich A, Schmedtje U. 1995. Indicating shear-stress with FST-hemispheres - effects
35 of stream-bottom topography and water depth. *Freshwater Biology* **34**: 107-
36 121.
37
38
39 Downes BJ. 1990. Patch dynamics and mobility of fauna in streams and other
40 habitats. *Oikos* **59**: 411-413.
41
42
43 Downes BJ, Lake PS, Glaister A, Bond NR. 2006. Effects of sand sedimentation on
44 the macroinvertebrate fauna of lowland streams: are the effects consistent?
45 *Freshwater Biology* **51**: 144-160.
46
47
48 Duncan MJ, Suren AM, Brown SLR. 1999. Assessment of streambed stability in
49 steep, bouldery streams: development of a new analytical technique. *Journal of*
50 *the North American Benthological Society* **18**: 445-456.
51
52
53 Effenberger M, Sailer G, Townsend CR, Matthaei CD. 2006. Local disturbance
54 history and habitat parameters influence the microdistribution of stream
55 invertebrates. *Freshwater Biology* **51**: 312-332.
56
57
58 Englund G. 1991. Effects of disturbance on stream moss and invertebrate community
59 structure. *Journal of the North American Benthological Society* **10**: 143-153.
60

- 1
2
3 Francoeur SN, Biggs BJE, Lowe RL. 1998. Microform bed clusters as refugia for
4 periphyton in a flood-prone headwater stream. *New Zealand Journal of Marine*
5 *and Freshwater Research* **32**: 363-374.
6
7
8
9 Graham AA, McCaughan DJ, McKee FS. 1988. Measurement of surface area of
10 stones. *Hydrobiologia* **157**: 85-87.
11
12 Holomuzki JR, Biggs BJE. 2000. Taxon-specific responses to high-flow disturbance
13 in streams: implications for population persistence. *Journal of the North*
14 *American Benthological Society* **19**: 670-679.
15
16
17 Hurlbert SH. 1971. The non-concept of species diversity: a critique and alternative
18 parameters. *Ecology* **52**: 577-586.
19
20
21 Keller EA, Melhorn WN. 1978. Rhythmic spacing and origin of pools and riffles.
22 *Geological Society of America Bulletin* **89**: 723-730.
23
24
25 Knighton D. 2008. *Fluvial forms and processes: a new perspective*. Arnold: London
26
27 Lake PS. 2000. Disturbance, patchiness, and diversity in streams. *Journal of the North*
28 *American Benthological Society* **19**: 573-592.
29
30
31 Matthaei CD, Huber H. 2002. Microform bed clusters: are they preferred habitats for
32 invertebrates in a flood-prone stream? *Freshwater Biology* **47**: 2174-2190.
33
34
35 Matthaei CD, Townsend CR. 2000. Long-term effects of local disturbance history on
36 mobile stream invertebrates. *Oecologia* **125**: 119-126.
37
38
39 McAuliffe JR. 1984. Competition for space, disturbance, and the structure of a
40 benthic stream community. *Ecology* **65**: 894-908.
41
42
43 McFarlane AG. 1951. Caddisfly larvae (Trichoptera) of the family Rhyacophilidae.
44 *Records of the Canterbury Museum* **5**: 267-289.
45
46
47 Merigoux S, Doledec S. 2004. Hydraulic requirements of stream communities: a case
48 study on invertebrates. *Freshwater Biology* **49**: 600-613.
49
50
51 Newbury RW. 1984. Hydrologic determinants of aquatic insect habitats. In: Resh VH,
52 Rosenberg DM (Editors). *The Ecology of Aquatic Insects*. Praeger Publishers,
53 New York, pp. 323-357.
54
55
56 Palmer MA, Bely AE, Berg KE. 1992. Response of invertebrates to lotic disturbance -
57 a test of the hyporheic refuge hypothesis. *Oecologia* **89**: 182-194.
58
59
60 Petts GE, Foster IDL. 1985. *Rivers and landscape*. Edward Arnold: London
Pfankuch DJ. 1975. Stream reach inventory and channel stability evaluation.,
U.S.D.A. Forest Service, Region 1, Missoula, Montana.

- 1
2
3 Poff NL, Ward JV. 1989. Implications of streamflow variability and predictability for
4 lotic community structure - a regional-analysis of streamflow patterns.
5 *Canadian Journal of Fisheries and Aquatic Sciences* **46**: 1805-1818.
6
7
8
9 Reice SR, Wissmar RC, Naiman RJ. 1990. Disturbance regimes, resilience, and
10 recovery of animal communities and habitats in lotic ecosystems.
11 *Environmental Management* **14**: 647-659.
12
13
14 Reid I, Frostick LE, Brayshaw AC. 1992. Microform roughness elements and the
15 selective entrainment of particles in gravel-bed rivers. In: Billi P, Hey RD,
16 Thorne CR, Tacconi P (Editors). *Dynamics of Gravel-Bed Rivers*. Wiley,
17 Chichester, pp. 253-275.
18
19
20
21 Resh VH, Brown AV, Covich AP, Gurtz ME, Li HW, Minshall GW, Reice SR,
22 Sheldon AL, Wallace JB, Wissmar RC. 1988. The role of disturbance in stream
23 ecology. *Journal of the North American Benthological Society* **7**: 433-455.
24
25
26 Riis T, Suren AM, Clausen B, Sand-Jensen K. 2008. Vegetation and flow regime in
27 lowland streams. *Freshwater Biology* **53**: 1531-1543. DOI: 10.1111/j.1365-
28 2427.2008.01987.x.
29
30
31
32 Sanders HL. 1968. Marine benthic diversity: a comparative study. *American*
33 *Naturalist* **102**: 243-282.
34
35
36 Schwendel AC, Death RG, Fuller IC. 2010. The assessment of shear stress and bed
37 stability in stream ecology. *Freshwater Biology* **55**: 261-281. DOI:
38 10.1111/j.1365-2427.2009.02293.x.
39
40
41 Schwendel AC, Death RG, Fuller IC, Joy MK. 2011a. Linking disturbance and stream
42 invertebrate communities - how best to measure bed stability. *Journal of the*
43 *North American Benthological Society* **30**: 11-24. DOI: 10.1899/09-172.1.
44
45
46 Schwendel AC, Fuller IC. in press. Connectivity in forested upland catchments and
47 associated channel dynamics: The Ruahine Range. *Journal of Hydrology (NZ)*.
48
49
50 Schwendel AC, Joy MK, Death RG, Fuller IC. 2011b. A macroinvertebrate index to
51 assess stream-bed stability. *Marine and Freshwater Research* **62**: 30-37. DOI:
52 10.1071/MF10137.
53
54
55 Steinman AD, Lamberti GA. 1996. Biomass and Pigments of Benthic Algae. In:
56 Hauer FR, Lamberti GA (Editors). *Methods in Stream Ecology*. Academic
57 Press, San Diego, pp. 295 - 313.
58
59
60

- 1
2
3 Suren AM. 1996. Bryophyte distribution patterns in relation to macro-, meso-, and
4 micro-scale variables in South Island, New Zealand streams. *New Zealand*
5 *Journal of Marine and Freshwater Research* **30**: 501-523.
6
7
8
9 Suren AM, Duncan MJ. 1999. Rolling stones and mosses: effect of substrate stability
10 on bryophyte communities in streams. *Journal of the North American*
11 *Benthological Society* **18**: 457-467.
12
13
14 Towns DR. 1981. Life histories of benthic invertebrates in a kauri forest stream in
15 Northern New Zealand. *Australian Journal of Marine and Freshwater Research*
16 **32**: 191-211.
17
18
19 Towns DR, Peters WL. 1996. *Leptophlebiidae (Insecta: Ephemeroptera)*. Fauna of
20 New Zealand, 36. Manaaki Whenua Press: Lincoln.
21
22
23 Townsend CR, Scarsbrook MR, Doledec S. 1997. Quantifying disturbance in streams:
24 alternative measures of disturbance in relation to macroinvertebrate species
25 traits and species richness. *Journal of the North American Benthological Society*
26 **16**: 531-544.
27
28
29
30 Vinson MR, Hawkins CP. 1996. Effects of sampling area and subsampling procedure
31 on comparisons of taxa richness among streams. *Journal of the North American*
32 *Benthological Society* **15**: 392-399.
33
34
35 Wild M, Snelder TH, Leathwick JR, Shankar U, Hurren H. 2005. Environmental
36 variables for the freshwater environments of New Zealand river classification,
37 NIWA, Christchurch.
38
39
40 Winterbourn MJ. 1973. A guide to the freshwater Mollusca of New Zealand.
41 *Tuatara* **20**: 141-159.
42
43
44 Winterbourn MJ. 2000. Stream communities and ecosystem processes. In: Collier KJ,
45 Winterbourn MJ (Editors). *New Zealand stream invertebrates: Ecology and*
46 *implications for management*. NZ Hydrological Society, Christchurch, pp. 13.1-
47 13.14.
48
49
50
51 Winterbourn MJ, Gregson KLD, Dolphin CH. 2006. *Guide to the aquatic insects of*
52 *New Zealand*. Bulletin of the Entomological Society of New Zealand 14.
53 Entomological Society of New Zealand: Auckland
54
55
56
57
58
59
60

Table I. Abiotic characteristics of the study sites assessed between October 2007 and March 2010. Depth, width, velocity, conductivity, temperature and pH measurements are averaged from 5 readings taken concomitant with invertebrate sampling, TTM is an index of bed stability calculated from the movement of *in situ* marked tracer stones. Sites used for validation are in italics.

site	Stream order (Strahler 1952)	Mean depth (m)	Mean width (m)	Mean flow velocity (m*s ⁻¹)	Mean conductivity (µS*cm ⁻¹)	Mean temperature (°C)	Mean pH	Substrate D ₅₀ (mm)	TTM
Tararua Range									
<i>Waitohu</i>	4	0.20	6.8	no data	84	11.5	7.5	86	39.94
<i>Waiotauru</i>	5	0.26	17.4	0.716	68	11.3	7.7	84	38.57
<i>Waikawa</i>	4	0.18	6.1	no data	76	13.2	7.5	99	36.78
<i>Panatewaewae</i>	3	0.11	7.0	no data	74	13.1	7.6	103	31.21
<i>Kiriwhakapapa</i>	3	0.15	6.5	0.603	64	9.7	7.2	59	20.43
<i>Ohau</i>	4	0.24	14.0	0.694	72	12.7	7.6	64	14.30
<i>Pukeatua</i>	3	0.18	9.7	0.720	80	12.4	7.7	84	12.50
<i>Makahika</i>	4	0.17	6.3	no data	66	19.4	7.4	82	11.18
<i>Mangatainoka</i>	4	0.17	11.7	0.694	52	13.3	7.1	108	7.67
<i>Rawnsley</i>	2	0.11	5.1	0.422	48	13.5	6.9	159	4.74
<i>Tokomaru</i>	4	0.14	14.6	0.707	81	17.5	6.9	85	3.07
<i>Kahuterawa</i>	4	0.15	3.5	0.616	68	14.1	6.5	85	0.06
Ruahine Range									
<i>Waipawa</i>	3	0.20	5.4	1.000	103	8.8	8.2	59	79.56
<i>Tamaki</i>	2	0.17	3.3	0.811	64	10.8	7.6	35	64.46
<i>Mangapuaka</i>	2	0.09	2.3	0.584	69	8.6	6.7	28	21.51
<i>Konewa</i>	3	0.10	6.5	0.575	133	13.0	7.5	72	15.36
<i>Rokaiwhana</i>	3	0.22	3.1	0.887	66	14.7	7.1	58	14.96
<i>Makawakawa</i>	4	0.21	27.6	0.630	58	14.3	7.0	83	12.03
<i>Raparapawai</i>	3	0.17	7.0	0.931	72	13.3	7.3	84	8.44
<i>Makiekie</i>	2	0.19	5.7	0.646	51	10.9	7.3	109	4.86
<i>Coppermine</i>	3	0.13	4.4	0.592	90	13.5	7.4	51	4.43
<i>Manawatu</i>	3	0.20	4.3	0.603	66	9.4	7.8	65	3.85
<i>Cone</i>	2	0.16	4.5	0.588	50	11.0	7.2	107	0.53
Central Plateau									
<i>Mangatoetoeuui</i>	4	0.26	8.6	0.597	139	7.1	8.0	97	18.70
<i>Waikato</i>	3	0.14	3.2	no data	66	10.0	7.8	18	11.38
<i>Te Piripiri</i>	3	0.19	2.0	0.742	69	8.5	7.7	35	2.09
<i>Wahianoa</i>	3	0.26	6.3	0.967	70	12.8	7.4	145	1.73
<i>Whakapapaiti</i>	4	0.28	15.8	0.976	138	11.8	8.2	125	1.02
<i>Oturere</i>	4	0.42	9.4	0.859	112	10.2	8.6	131	0.59
<i>Makomiko</i>	3	0.13	5.3	no data	27	11.3	7.5	107	0.07
<i>Makotuku</i>	2	0.13	5.7	no data	30	9.2	7.4	116	0.03
<i>Waiharakeke</i>	3	0.23	3.5	0.965	159	10.7	8.1	104	0.01
<i>Mangahuia</i>	2	0.20	8.6	no data	38	9.9	7.1	147	0.01
<i>Poutu</i>	5	0.44	7.7	1.053	71	10.6	8.0	80	0.00
<i>Orautoha</i>	3	0.24	2.5	0.548	128	12.8	8.3	166	0.00
<i>Pukeonake</i>	4	0.17	6.8	0.398	23	8.2	7.0	158	0.00
Mt. Egmont									
<i>Waiwhakaihoh</i>	3	0.25	13.8	0.614	109	16.6	7.9	100	50.00
<i>Timaru</i>	2	0.12	3.6	0.295	69	14.6	6.9	213	34.04
<i>Kaiauaui</i>	3	0.17	11.8	0.756	159	17.5	7.9	172	26.68
<i>Manganui</i>	2	0.15	14.7	1.022	56	18.9	6.7	142	20.77

1										
2										
3										
4										
5										
6										
7										
8										
9										
10										
11										
12										
13										
14										
15										
16										
17										
18										
19										
20										
21										
22										
23										
24										
25										
26										
27										
28										
29										
30										
31										
32										
33										
34										
35										
36										
37										
38										
39										
40										
41										
42										
43										
44										
45										
46										
47										
48										
49										
50										
51										
52										
53										
54										
55										
56										
57										
58										
59										
60										

For Peer Review

Table II. Assessed properties of the channel, banks and riparian environment potentially related to bed stability, categorical variables were rated at a scale from 1 (associated with stable substrate) to 4 (associated with substrate instability), * variable removed because of intercorrelation.

variable	description
Riparian environment	
A11	Fraction of pasture on riparian strip (%)
A12	Fraction of native forest on riparian strip (%)
A13	Fraction of exotic vegetation on riparian strip (%)
A14	Fraction of scrub on riparian strip (%)
A15	Fraction of other land cover (none, tussock, etc.) on riparian strip (%)
A21	Ratio of floodplain width to active channel width (m/m)
A22*	Ratio of floodplain width to wet channel width (m/m)
A23	Ratio of active channel width to wet channel width (m/m)
A31	Percentage of high bank surface covered with vegetation (%)
A32	Variation in species and age of high bank vegetation (categorical)
Channel cross profile	
B11	Channel incision, ratio of width to depth (m/m)
B21	Bank erosion (categorical)
B22	Number of recent bank collapses
B31	Number of recently deposited lateral bars of fine material (< coarse gravel)
Channel longitudinal profile	
C11	Bed slope (m/m)
C21	Sinuosity (categorical)
Channel bottom	
D11	Fraction of area affected by erosion and deposition (%)
D21	Occurrence and form of aquatic vegetation (categorical)
D31	Number of multiple barforms
D32	Fraction of area occupied by multiple barforms (%)
D41	Number of riffle-pool and step-pool sequences
D51	Occurrence of bedform clusters (categorical)
D61	Fraction of area with supercritical flow (%)
Substrate	
E11	Grain angularity (categorical)
E21	Constitution of grain surface (categorical)
E31	Interlock and overlap between particles (categorical)
E41	Packing and compaction of particles (categorical)
E51	Fraction of sand and smaller grain size (% area)
E52	Fraction of gravels (% area)
E53	Fraction of cobbles (% area)
E54	Fraction of boulders (% area)
E55	Homogeneity (% area of most abundant size class/ number of size classes present)
E56*	Size index (Sum of fractions weighted by their geometrical mean size of their size class)
E57*	Mean size index (Size index/ number of size classes present)
E58*	Fraction of cobbles and gravels (% area)
E61	Fraction of stable material (large boulders and bedrock) (%)
E71	Occurrence of an armour layer (categorical)

Table III. Correlation of bed stability measurements (total tracer movement – TTM, bottom component Pfankuch Index – BCP), measured (marked with *) environmental parameters and periphyton biomass and downstream variables, segment variables and runoff-weighted upstream catchment variables from the FWENZ database (Wild *et al.*, 2005) with NMDS axes, significant correlations are marked bold ($p < 0.01$).

Axis	1	2
	Pearson's R	Pearson's R
Width*	-0.06	0.06
Depth*	0.33	-0.20
Velocity*	0.12	-0.19
Conductivity*	0.22	-0.03
Temperature*	-0.17	0.13
pH*	0.33	-0.09
Riparian Pasture*	0.18	0.25
Riparian bare floodplain*	-0.44	-0.33
Periphyton biomass*	0.44	0.13
Average slope of <i>downstream</i> network	-0.31	0.09
Maximum slope of <i>downstream</i> segments	0.18	-0.10
Maximum <i>segment</i> slope based on 30 m grid	0.03	0.17
<i>Segment</i> sinuosity	-0.05	0.07
Average <i>segment</i> slope	-0.26	-0.12
Shaded fraction of <i>segment</i>	-0.02	0.06
Percentage of the <i>segment</i> riparian area covered in scrub	0.20	0.13
<i>Upstream</i> mean January air temperature	0.06	0.49
<i>Upstream</i> catchment rain days > 15 mm/month	-0.27	0.13
<i>Upstream</i> lake index	0.19	0.00
Percentage of <i>upstream</i> catchment annual runoff from alluvium	0.12	0.13
Percentage of <i>upstream</i> catchment annual runoff from peat	-0.12	-0.03
<i>Upstream</i> average of calciferous regolith	-0.19	0.14
<i>Upstream</i> catchment average of regolith hardness	-0.10	-0.06
<i>Upstream</i> catchment average of particle size	-0.05	0.04
Percentage of <i>upstream</i> catchment consists of bare ground	0.15	-0.62
Percentage of <i>upstream</i> catchment covered in exotic forest	0.25	-0.10
Percentage of <i>upstream</i> catchment covered in indigenous forest	-0.08	0.39
Percentage of <i>upstream</i> catchment with pastoral landuse	0.17	0.09
Percentage of <i>upstream</i> catchment covered in tussock	-0.04	-0.25
Percentage of <i>upstream</i> catchment consist of wetland	0.10	0.10
<i>Segment</i> stream order	0.18	-0.02
TTM	-0.53	0.04
BCP	-0.57	0.09

Table IV. Results of the regression analysis of the NMDS axis against 39 characteristics of the channel and the riparian environment ($R^2 = 0.805$, adjusted $R^2 = 0.726$), VIF – variance inflation factor.

Variables	Coefficient	Std error	T-test if slope $\neq 0$	P value	VIF
Constant	-6.31006	1.00297	-6.29	0	0
A23	0.21652	0.06028	3.59	0.0011	2.1
A31	0.01239	0.00375	3.31	0.0023	1.8
B21	0.26123	0.06495	4.02	0.0003	2
C11	0.05583	0.02096	2.66	0.012	1.3
D11	0.29004	0.09619	3.02	0.005	3.2
D31c	0.28711	0.07222	3.98	0.0004	3
D32	0.012	0.00556	2.16	0.0385	1.9
D51c	0.27049	0.07771	3.48	0.0015	1.6
E11	0.2418	0.12253	1.97	0.0572	1.5
E21	0.16677	0.09457	1.76	0.0874	2.6
E41	0.25041	0.11964	2.09	0.0444	1.7
E51	0.02885	0.00937	3.08	0.0042	2.2
E55	0.0524	0.02019	2.6	0.0141	3.1

Table V. Correlation of invertebrate community metrics with bed stability assessed with the SBSI protocol, *in situ* marked tracer stones (TTM) and the bottom component of the Pfankuch Stability Index (BCP) at 46 New Zealand streams used for SBSI calibration and at 8 independent sites from the same regions for validation. Significance from multiple correlations was adjusted using False Discovery Rate and is indicated by * for $\alpha = 0.05$, ** for $\alpha = 0.005$ and *** $\alpha = 0.001$.

	SBSI calibration sites			Validation sites		
	SBSI	TTM	BCP	SBSI	TTM	BCP
Brillouin Index	-0.75***	-0.52***	-0.68***	-0.81*	-0.78*	-0.73*
Taxa number	-0.56***	-0.27	-0.54***	-0.73*	-0.34	-0.82*
Rarefied taxa number for 200 individuals	-0.77***	-0.51***	-0.55***	-0.73*	-0.40	-0.82*
Mean number of individuals	-0.75***	-0.35*	-0.45**	-0.74*	-0.34	-0.86*

Table VI. Correlation of bed stability assessed with the SBSI protocol, *in situ* marked tracer stones (TTM) and the bottom component of the Pfankuch Stability Index (BCP) at 46 New Zealand streams used for SBSI calibration and at 8 independent sites from the same regions for validation. Significance from multiple correlations was adjusted using False Discovery Rate and is indicated by * for $\alpha = 0.05$, ** for $\alpha = 0.005$ and *** $\alpha = 0.001$.

	SBSI calibration sites			Validation sites	
	TTM	BCP		TTM	BCP
SBSI	0.48***	0.66***		0.47	0.75*
BCP	0.46**			0.67	

Table VII. Parameters of the stream Bed Stability for Invertebrates (SBSI) survey with weights and potential range of values (*extreme values estimated) and scores.

	Parameter	Weight	Range	Minimum score	Maximum score
C11	Bed slope	0.56	0.0001* – 1*	0.00006	0.56
A23	Active /wet channel	2.17	1 – 10*	2.17	21.7
B21	Bank erosion	2.61	1 – 4	2.61	10.44
A31	Bank vegetation cover	0.12	0 – 100	0	12
E51	Sand fraction	0.29	0 – 100	0	29
E55	Substrate homogeneity	0.52	4 – 100	2.08	52
E41	Packing and compaction	2.50	1 – 4	2.50	10.0
E21	Particle surface	1.67	1 – 4	1.67	6.68
E11	Grain angularity	2.42	1 – 4	2.42	9.68
D11	Reworked area	2.90	1 – 4	2.90	11.60
D31	Multiple barform number	2.87	0 – 5	0	14.35
D32	Area of multiple barforms	0.12	0 – 100	0	12
D51	Bedform clusters	2.70	1 – 4	2.70	10.80
	Total SBSI			19.05	200.81

1
2
3
4 Figure 1. Stream reaches in the southern North Island of New Zealand studied for
5 calibration of the Stream Bed Stability for Invertebrates protocol. Open circles denote
6 the sites used for validation.
7
8
9

10
11 Figure 2. Non-metric Multidimensional Scaling axes of 46 mountain stream
12 invertebrate communities and correlated parameters ($p < 0.01$). Periphyt – periphyton
13 biomass, usAveTWar - Upstream mean January air temperature, usIndigF -
14 Percentage of upstream catchment covered in indigenous forest, TTM – total tracer
15 movement, BCP – bottom component of Pfankuch Index, RipBareF - Dry active
16 channel bare of vegetation under base flow conditions, usBare_q - Percentage of
17 upstream catchment consisting of bare ground.
18
19
20
21
22
23

24
25 Figure 3. Stream bed stability assessed with the Stream Bed Stability for Invertebrates
26 Index (SBSI), *in situ* marked tracer stones (TTM) and the bottom component of the
27 Pfankuch Stability Index (BCP) plotted against the NMDS axis used for calibration of
28 the SBSI.
29
30
31
32

33
34 Figure 4. Site scores of the Stream Bed Stability for Invertebrates Index (SBSI)
35 plotted against conventional measures of bed stability: *In situ* marked tracer stones
36 (TTM, closed symbols) and the bottom component of the Pfankuch Stability Index
37 (BCP, open symbols). Sites used for validation are shown as triangles.
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

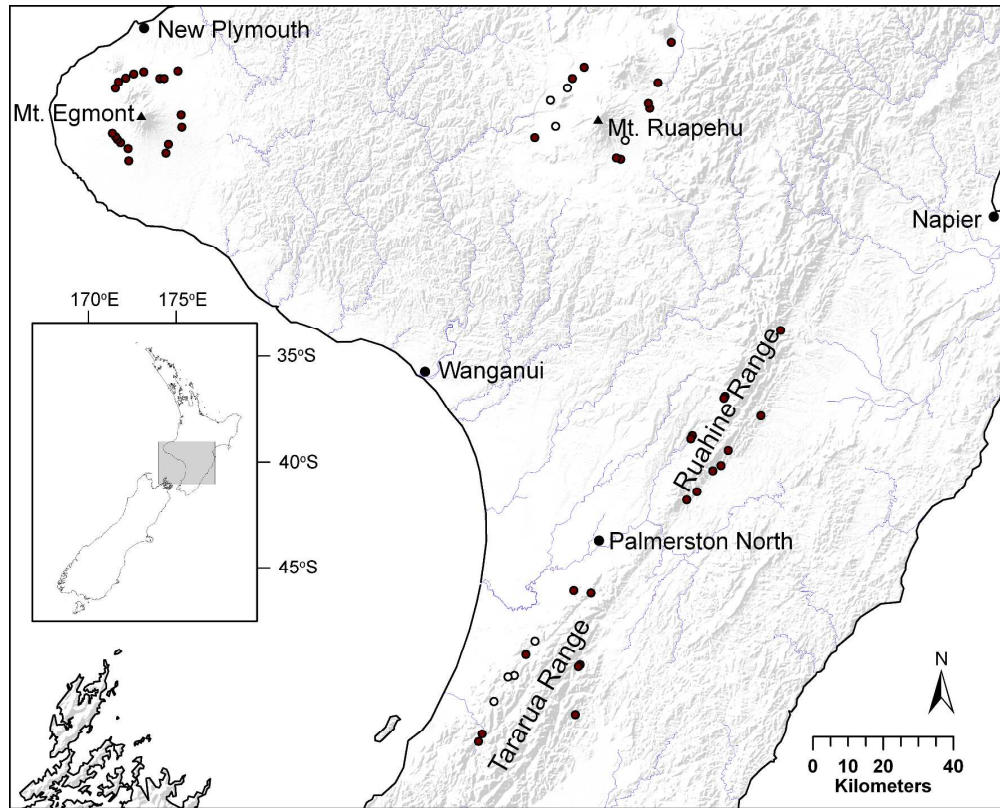


Figure 1. Stream reaches in the southern North Island of New Zealand studied for calibration of the Stream Bed Stability for Invertebrates protocol. Open circles denote the sites used for validation.
150x120mm (600 x 600 DPI)

view

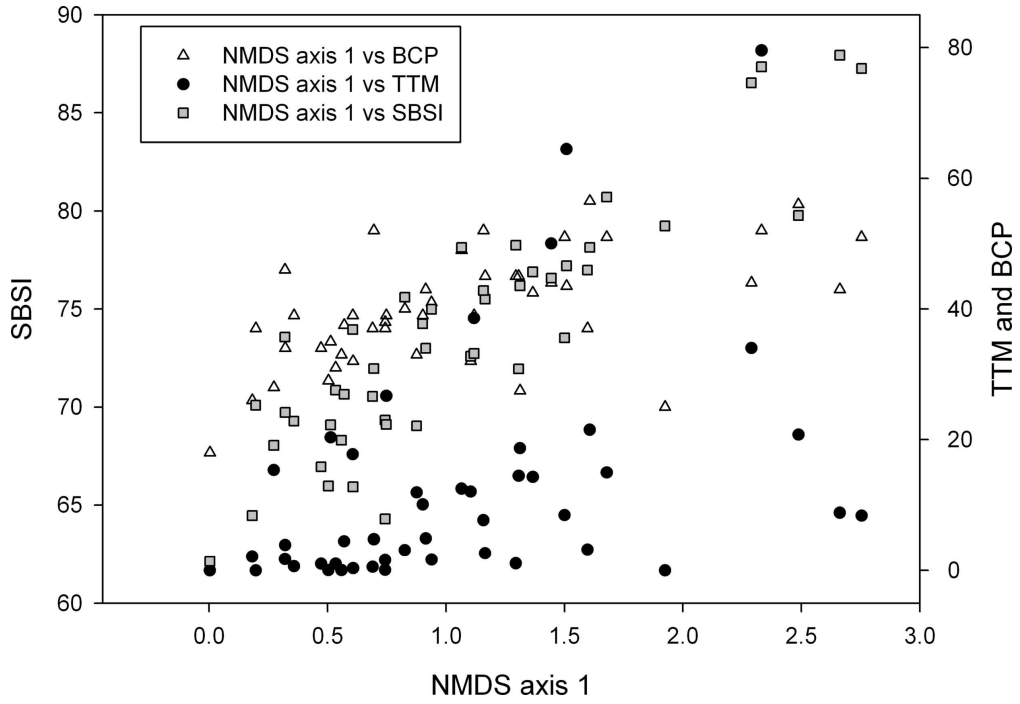


Figure 3 Stream bed stability assessed with the Stream Bed Stability for Invertebrates Index (SBSI), in situ marked tracer stones (TTM) and the bottom component of the Pfankuch Stability Index (BCP) plotted against the NMDS axis used for calibration of the SBSI.
105x72mm (600 x 600 DPI)

Review

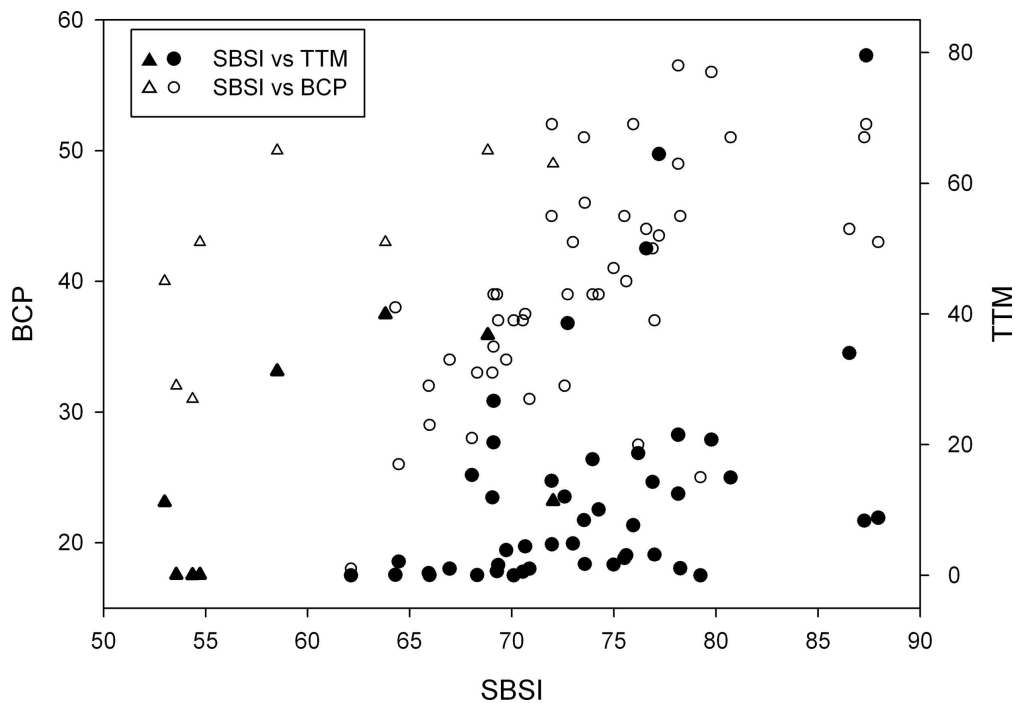


Figure 4 Site scores of the Stream Bed Stability for Invertebrates Index (SBSI) plotted against conventional measures of bed stability: In situ marked tracer stones (TTM, closed symbols) and the bottom component of the Pfankuch Stability Index (BCP, open symbols). Sites used for validation are shown as triangles.
105x72mm (600 x 600 DPI)

review