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Reduced inorganic sulfur in surface sediment and its impact on
benthic environments in offshore areas of NE China
Yanqing Sheng ^{1, *} , Qiyao Sun ¹ , Simon Bottrell ² , Robert Mortimer ³
¹ Research Center for Coastal Environmental Engineering Technology of Shandong
Province, Yantai Institute of Coastal Zone Research, Chinese Academy of Sciences,
Yantai 264003, P.R. China
² School of Earth and Environment, University of Leeds, Leeds LS2 9JT, UK
³ School of Animal, Rural and Environmental Sciences, Nottingham Trent University,
Brackenhurst campus, Southwell, Nottinghamshire, NG25 0QF, UK
Abstract
Geochemical cycling and biological toxicity of sulfur in marine sediments is
closely related to the activity of organisms. This study investigated the distribution
and potential impact on benthic environments of acid volatile sulfur (AVS),
chromium (II)-reducible sulfur (CRS), elemental sulfur (ES), total S, C, N and Fe in
superficial sediments across the Bohai Sea, Yellow Sea and East China Sea. The
composition of reduced inorganic sulfur in the three study areas was dominated by
CRS (averaging 72% of total reduced inorganic sulfur). The low AVS content
*Corresponding author: Tel.: +86 535 2109265; Fax: +86 535 2109000; E-mail address:

yqsheng@yic.ac.cn

(average of 1.12 µmol g⁻¹) of the sediments and the low values of AVS/CRS (average
0.34 µmol g⁻¹), degree of pyritization and degree of sulphidization indicate that there
is sufficient available iron in the sediment to restrict the threat of sulphide toxicity to
benthic organisms in most of the study areas. However, high organic matter loads in
parts of the study areas have resulted in enhanced accumulation of AVS, resulting in a
higher toxicity risk.

7

8 Keywords: Sulfur compounds; Sedimentary environments; Offshore; Benthic
9 environment

10 1. Introduction

11 Sulfur is essential to life and one of the key players in global biogeochemical cycles. 12 However the most reduced form, sulfide, is generally toxic to organisms and influences water quality (Bagarinao, 1992; Sheng et al., 2011). Sulfur is cycled 13 between and fixed in many different forms in marine sediments (Morse and Berner, 14 15 1995; Canfield et al., 2005; Bottrell et al., 2009; Alvarez and Rubio, 2012). Therefore, understanding sulfur speciation in marine sediments is essential in evaluating its 16 17 response to environmental change and the potential biological impact of sulfide. The 18 availability of sulfide in the coastal environment will have implications for overlying 19 water quality.

The sulfur cycle in marine sediments can be divided into reductive and oxidative processes. The reductive side of the cycle is driven by microorganisms that reduce

2

sulfate to sulfide in anaerobic environments and rates of sulfate reduction can be very high under such conditions (Morse et al., 2007; Tarpgaard et al., 2011).

3
$$SO_4^{2-} + 2CH_2O => H_2S + 2HCO_3^{-}$$
 (1)

In the oxidative side of the cycle sulfide reacts with a variety of oxidants such as O_2 4 and Fe(III) oxides to produce sulfate or intermediate S forms (e.g. elemental S). The 5 majority of the sulfide produced in marine sediment is reoxidized (Canfield and 6 Thamdrup 1994; Poulton 2003; Bottrell and Newton, 2006). Sulfide produced is also 7 removed by reaction with mineral-derived Fe, initially producing a monosulfide 8 "FeS" which transforms to more stable pyrite, FeS₂ (Berner 1970, 1984). However, 9 the reactivity of mineral iron forms is highly variable (Canfield et al. 1992) and may 10 11 limit the removal of sulfide. Where supply of oxidants is also restricted, production of 12 sulfide may outcompete rates of removal and concentrations of toxic sulfide will increase in shallow pore-waters and bottom waters (Phillips et al. 1997). Furthermore, 13 14 iron monosulfide species formed in sediment are often highly reactive and will rapidly react with and remove dissolved oxygen from bottom waters if sediment is 15 resuspended. 16

The reactivity of sulfide stored in sediment when resuspension occurs is dependent on the form in which sulfide is present; dissolved sulfide is most reactive, followed by monosulfide, and pyrite is least reactive (Morse and Rickard 2004). In many sediments conversion of monosulfide to pyrite is rapid (Schoonen and Barnes 1991; Butler and Rickard 2005) such as reoxidation of AVS in the uppermost layer

and thus monosulfide concentrations are low or negligible (e.g. Rozan et al. 2002; 1 Rickard and Morse 2005). However in other situations (e.g. high pH (>7.8) sediments 2 3 and under organic-rich eutrophic conditions) Fe monosulfides are persistent and represent a significant pool of reactive sulfide in the sediment (Hurtgen et al. 1999; 4 Morse 1999; Burton et al. 2011; Morgan et al., 2012). Sulfide is toxic to aquatic plants 5 and organisms. Its accumulation in upper sediment layers in coastal areas exerts an 6 impact upon not only local benthos but also the pelagic biota due to the potential 7 release of free sulfide into the water column (Bagarinao, 1992). Furthermore, 8 9 re-suspension of sediment can result in oxidation of H₂S and reactive sulfides, changing the redox conditions of the overlying water and leading to hypoxia at the sea 10 11 floor (Phillips et al., 1997; Sorokin and Zakuskina, 2012). Oxidation of monosulfide 12 in surface sediments may also lead to the release of toxic heavy metals (coprecipitated with FeS) into more bioavailable forms, such as dissolved Cu and Zn (Simpson et al., 13 2012; Rodrigues et al., 2013). Most organisms live within oxic surface sediments or in 14 oxygenated microenvironments created within sub-oxic and anoxic sediments 15 16 (Simpson et al., 2012), and hence are vulnerable to shifts in redox and associated 17 changes in sulfur speciation and toxicity.

18 The degree of pyritization (DOP) and the degree of sulphidization (DOS) are 19 two parameters that can be used to distinguish between situations where pyrite and Fe 20 sulfide formation is either C- or Fe-limited and to measure the completeness of the 21 reaction of reactive Fe (operationally defined as the fraction of Fe that is soluble in 1 22 M HCl for 16h) with aqueous sulfide (Raiswell and Berner, 1985). In sediments that

contain a significant amount of Fe monosulfide, DOP underestimates the amount of 1 Fe that has reacted with H₂S (Boesen and Postma, 1988). Instead, DOS represents the 2 3 degree to which reactive Fe has been transformed into sulfide and, therefore, provides a better indication of Fe limiting conditions (Yin et al., 2008). In sediments where Fe 4 availability becomes limiting pore-water sulfide concentrations are likely to increase, 5 enhancing the likelihood of sulfide toxicity. Conversely, if sulfide production is 6 7 limited by the availability of a labile organic substrate, pore-water sulfide concentrations may be very low. 8

9 The Bohai Sea, Yellow Sea and East China Sea are situated north-east (NE) of China. They are important production bases for fishery and marine industries. This 10 11 study area can be characterized as a region surrounded by areas of high population 12 growth and economic development. A large amount of pollutants are discharged into these areas in industrial and municipal wastewaters via rivers (i.e. Yellow River, 13 Yangtze River and other smaller rivers), along with excess inputs of nutrients from 14 15 mariculture. This pollution may influence the cycles of iron, sulfur and phosphorus 16 (Rozan et al., 2002), leading to offshore eutrophication and benthic macroalgal 17 blooms, especially large offshore phytoplankton blooms e.g. enteromotpha prolifera in Yellow Sea (Liu et al., 2013). Previous studies on the effects of biogenic elements 18 19 in sediments in these areas were focused mainly on carbon, nitrogen and phosphorus (Liu et al., 2004; Liu and Yin, 2007). However, to date, only limited effort has been 20 21 devoted to the study of sedimentary geochemistry of reduced inorganic sulfur (RIS) in the offshore areas of NE China (Kang et al., 2014), and its potential impact on the 22

1 marine ecosystem remains unclear.

In this study, acid volatile sulfur (AVS), chromium (II)-reducible sulfur (CRS, pyritic sulfur), elemental sulfur (ES), total sulfur (TS), reactive iron, total organic carbon (TOC) and total nitrogen (TN) were analyzed in surface sediments from the study area. The partitioning of RIS was used to understand sulfur biogeochemistry within the sediments, and to predict the potential for ecotoxicological risk due to release of toxic sulfide.

8

9 **2. Methods and materials**

10 2.1 Samples collection and handling

11 A total of 82 sediment samples were collected from the Bohai Sea, Yellow Sea 12 and East China Sea (Fig. 1). Surface sediments (0-20 cm) were collected using a stainless steel grab sampler and were immediately isolated from the atmosphere in 13 plastic zip-lock bags with all air expelled by N2 gas (a N2 gas cylinder was aboard 14 15 ship). Samples were frozen and stored in an icebox (-20°C) until arrival at the laboratory. Samples were processed immediately upon return to the laboratory. In the 16 17 laboratory, the plastic sample bags were first flushed with high purity N₂ again. Samples were then homogenized under this inert atmosphere (a glove box) by mixing 18 with a plastic spatula. Four replicates were used throughout. 19

- 20
- 21

Fig. 1

1 2.2 Sample analysis

The reagents used were all analytical grade or above, and deionized water 2 3 (milli-Q) was used to prepare reagent solutions. All glass and plastic were soaked in 10% HNO₃ for 48 h and rinsed with milli-Q water three times before use. Total 4 organic carbon (TOC) in sediments determined by a Shimadzu 5 was TOC-V_{CPH}/SSM-5000A. Total nitrogen (TN) and total sulfur (TS) were determined 6 by an Elementar vario MACRO cube CHNS analyzer. The precision of the 7 measurements was within 5% based on three replicate sediment analyses. Prior to 8 9 grain size analysis, each sediment sample was treated with sodium hypochlorite (NaOCl) to remove organic matter. Sample granulometry was analyzed using a 10 11 Malvern Mastersizer 2000 laser diffractometer capable of analyzing particle sizes 12 between 0.02 and 2,000 µm. The percentages of samples in each of the following three grain-size groups were determined: $< 4 \mu m$ (clay), 4-63 μm (silt), and $>63 \mu m$ 13 14 (sand).

15 The separation and determination of AVS, CRS and ES were conducted using a modified procedure described by Newton et al. (1995) and Hsieh & Shieh (1997). 16 17 Reagents and extraction steps of sulfur species separation are the same as the description of Hsieh & Shieh (1997), the detection of H₂S is the same as the 18 19 description of Newton et al. (1995). In this method, a thin glass pipe (with N₂ gas inside) was dipped into corresponding solutions (shaking occasionally) to accelerate 20 21 the emission of H₂S (1.5 h). All subsequent processing of sediment was performed under a N₂ atmosphere inside a glove box at room temperature. Briefly, AVS, CRS 22

1	and ES were separated sequentially by 6 M HCl, acidic Cr (II) and Cr (II) plus N,
2	N-dimethylformamide, respectively, using pure N_2 as carrier gas to purge and trap
3	H ₂ S, at a temperature of ~ 60 $^{\circ}$ C (by electric hot plate). The liberated H ₂ S was trapped
4	in a 0.1 mol L^{-1} CuCl ₂ solution. The amount of H ₂ S evolved from the sample for each
5	solid-phase RIS species was determined by titrating the Cu remaining in solution with
6	0.1 mol L ⁻¹ EDTA (assuming a precipitate of CuS stoichiometry). Three to four drops
7	of glycine cresol red (0.5% aqueous) was used as an indicator and the titration was
8	buffered with 70 ml of 1 mol L^{-1} sodium acetate solution (adjusted to pH 5.5 with
9	acetic acid). The endpoint was sharp and marked by a colour change from dark blue to
10	a light green. Sulfide solutions (Na ₂ S) with concentrations of 0.5 and 1 mg L^{-1} were
11	used in recovery experiments. Average sulfide recovery rate was 96%. Measured
12	sulfide data were converted to real sulfide contents after correction for the recovery
13	rate. The precision of triplicate analysis was within 5% for different fractions of RIS.
14	The fractions of Fe were determined by sequential acid digestion. Reactive Fe is the
15	fraction of Fe that is soluble in 1 mol L^{-1} HCl (16h, Fe _R) and pyrite Fe (Fe _{py}) is the Fe
16	that is soluble in concentrated HNO ₃ , after removal of the Fe associated with silicates
17	and organic matter (Huerta-Dı'az & Morse, 1990). The concentrations of different Fe
18	fractions (extracted by corresponding acid) were detected by an Atomic Absorption
19	Spectrometer (TAS 990, Beijing Purkinje General Instrument Co.Ltd., Beijing),
20	precision was to within 2% based on triplicate analysis.

	DOP was taken to be the percentage of pyrite-Fe in the bioavailable iron, which
2	was calculated as $Fe_{py}/(Fe_R + Fe_{py})$ (Berner, 1970). Fe_{py} was calculated from FeS_2
3	stoichiometry. DOS was taken to be the percentage of Fe that has been transformed
4	into sulfide, which was defined by $(Fe_{py} + Fe_{AVS})/(Fe_{py} + Fe_R)$ (Boesen and
5	Postma,1988). Fe _{AVS} is sulfide-bound Fe(II) assuming that AVS predominantly
6	occurs as FeS, though this assumption remains controversial (Morse and Rickard,
7	2004; Rickard and Morse, 2005).
8	
9	3. Results and discussion
10	3.1 Distribution of grain size
11	
12	Fig. 2
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-	
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located in the main water exchange channel between the Bohai Sea and the Yellow 1 Sea (Liu et al., 2004), so the small particles (clay) may be winnowed by strong water 2 3 currents, resulting in a predominance of sand in this area (Fig.2). In the East China Sea, driven by the Zhejiang-Fujian coastal current, a large amount of fine grained 4 particulates from the Yangtze Estuary are transported southward along the coast and 5 trapped in the inner shelf by the blocking of the northward warm Taiwan current 6 offshore, developing mud wedges outside the Yangtze River mouth (Xu et al., 2009). 7 Therefore, at site E01 (Fig. 1), the sediment was dominated by fine grained 8 9 particulates, with silt and clay accounting for ~ 97% of total grains (Fig. 2).

10

11 3.2 Distribution of TOC, TN and TS

12 The surface sediments were characterized by variable concentrations of TN, TOC and TS. TN varied from 0.03-0.21% (% dry weight of the sediment), TOC from 13 0.11- 2.49% and TS from 0.11-0.40%. The average TN in the Bohai Sea, Yellow Sea 14 15 and East China Sea were 0.07%, 0.08% and 0.06%, respectively. The averages for TOC were 1.23%, 1.07% and 1.09% respectively (Fig. 3). The highest TOC contents 16 17 were recorded at site B66 (2.3%) in the Bohai Sea, B31 (2.5%) in the Yellow Sea and E01 (1.6%) in the East China Sea. In the Bohai Sea, TN, TOC and TS exhibited high 18 values in Bohai Bay and Laizhou Bay. In the Yellow Sea, the TOC and TS contents in 19 sediments at B31 and B32 were high (2.5% and 1.5% respectively), indicating high 20 21 organic pollution loads in these areas, most likely related to the Dalian crude oil spill in 2010 (Lv et al., 2011). This organic pollution may provide an additional substrate 22

for sulfate reduction and lead to increased sulfide production and accumulation in the
 sediment.

- 3
- 4

5

Fig. 3

6 3.3 Distribution characteristics of different fractions of RIS

7 Fig. 4 presents the AVS, CRS and ES concentrations in the surface sediment samples of the Bohai Sea, Yellow Sea and East China Sea. The AVS, CRS, and ES 8 values were in the ranges $0.2-4.1\mu$ mol g⁻¹, $0.6-101\mu$ mol g⁻¹, and $0.5-3.9\mu$ mol g⁻¹, 9 respectively. AVS contents in the Bohai Sea, Yellow Sea and East China Sea are 10 close to the ranges of AVS contents for Jiaozhou Bay sediments $(0.06-13.7 \mu mol g^{-1})$ 11 (Zhu et al., 2012) and East China Sea shelf sediments $(0-25\mu mol g^{-1})$ (Lin et al., 2000; 12 Zhu et al., 2013). For CRS, the concentrations are 0.6-101µmol g⁻¹, dominating the 13 total RIS in all studied areas, with an average 75% share in the Bohai Sea, 59% in the 14 15 Yellow Sea and 87% in the East China Sea (Fig. 4 and Table 1). This finding was consistent with the reports of Zhu et al. (2012) in Jiaozhou Bay (CRS: 8.7-51.1µmol 16 g⁻¹), Sheng et al. (2013) in the coastal zone of the Yellow Sea (CRS: 5.5-13.1µmol g⁻¹) 17 and Zhu et al. (2013) in the East China Sea inner shelf (CRS: 2.7-38.3µmol g⁻¹). In 18 19 this study, although there are some values of RIS at sites B30, B31 and B32 associated with an oil spill, even these highest values (101.2 μ mol g⁻¹) are much lower than the 20 peak value for the southern East China Sea slope sediments (range: 0-240 μ mol g⁻¹) 21 22 (Lin et al., 2002). CRS dominates over AVS in the RIS pool in the Bohai Sea, Yellow

Sea and East China Sea sediments and thus in all these locations AVS produced by 1 sulfate reduction has been efficiently transformed to pyrite and there is limited 2 3 accumulation of reactive monosulfide. Because of high concentrations of reactive iron (average is approximately 100 μ mol g⁻¹) in these areas, the sulfide may be removed 4 quickly by the formation of FeS, then it was transformed to FeS₂ due to sufficient ES 5 $(0.5-3.9 \ \mu mol g^{-1})$ is present under anoxic conditions (reoxidation of AVS would be 6 7 restrained because of limited free oxygen in benthic bottom), resulting in high concentration of CRS. Overall, accumulation concentrations of labile sulfides (i.e. 8 9 AVS) in the study area is much lower than levels known to be potentially toxic for aquatic biota (200 mg S L⁻¹ of wet silt, Sorokin and Zakuskina, 2012), suggesting the 10 11 environmental conditions are low risk for the local marine ecosystem.

12 Generally, RIS concentrations were higher in the Yellow Sea than in the Bohai Sea and East China Sea, especially at sites B30, B31 and B32. This phenomenon is 13 consistent with the variation of TOC and TS in same sites, and is most likely related 14 15 to the Dalian crude oil spill summer 2010 (Lv et al., 2011). Deposited crude oil may 16 result in a high load of organic pollutants in these areas, increasing TOC accumulation 17 in the sediment (B30, B31 and B32). Furthermore, these organic compounds may provide an additional substrate for sulfate reduction and lead to increased sulfide 18 production and accumulation in the sediment. Whilst the oil spill has modified the 19 sediment geochemistry and increased AVS content, in the samples analyzed this 20 21 increase is small and lies within the range of many sediments that have not been polluted by oil spills. 22

2

Fig. 4

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4 3.4 Relationships between C, N, S and different fractions of RIS

5 The atomic molar ratios of C/N, C/S and AVS/CRS are listed in Table 1. C/N and C/S are ratios of TOC concentrations against TN and TS concentrations, respectively. 6 7 C/N is widely applied in biogeochemical studies of terrestrial, riverine, estuarine, and marine environments, most commonly to estimate the fraction of terrestrially derived 8 9 organic matter in marine sedimentary samples (Meyers, 1997). In this study, the C/N ratio (molar ratio) varied from 10.1 to 48.4 (average: 18.9) in the Bohai Sea, 7.7-41.3 10 11 (average: 15.1) in the Yellow Sea and 16.5-20.7 (average: 17.8) in the East China Sea 12 (Table 1). These results suggest that the studied areas are abundant in terrestrial-derived organic matter with high C/N ratios (always higher than 10) 13 14 (Meyers, 1997), associated with material rich in cellulose which originates from 15 terrestrial vascular plants and reaches the sediments via rivers, i.e. Yellow River and 16 Yangtze River.

TS and TOC/TS (C/S) are used for studying oxic/anoxic conditions in marine environments (Berner, 1984). The C/S ratios varied from 6.1 to 37.5, with an average of 12.8 in the Bohai Sea, 1.3-24.2 (average: 10.3) in the Yellow Sea and 6.5-17.0 (average: 10.3) in the East China Sea. Generally, the ratio of C/S in normal marine sediment (deposited under an oxic water column) is 2.0-3.8 (Berner, 1982). Overall, the C/S ratio in this study was highly variable at 1.3-37.5 (average: 11.4), with values

1	in most sampling sites higher than is normal for marine sediments. This can be
2	explained by the impact of freshwater input and mariculture activity, which can both
3	significantly influence C/S ratios, e.g. site B66 (37.5), is close to the Yellow River
4	estuary (with high TOC and low RIS values in sediment); site H1 (24.3) is close to
5	Qingdao city, where there are many large wastewater treatment plants (over 1 million
6	tons discharged per day); site H35 (21.7) is in a mariculture area. Generally, sewage
7	and mariculture contribute more C but not more S, resulting in high C/S ratios. The
8	result indicates that sulfate reduction or RIS accumulation was variable in surface
9	sediment of different areas, and may be influenced by anthropogenetic activities
10	(organic matter input) and redox conditions. AVS/CRS varied from 0.14 to 1.22, with
11	an average of 0.48 in Bohai Sea, 0.01-1.09 (average: 0.22) in Yellow Sea and
12	0.02-0.11 (average: 0.06) in East China Sea. Low ratios indicate that the sediment in
13	these areas poses a low threat of toxicity to benthic organisms because CRS is more
14	stable and less likely to release sulfide or metals than AVS, which is more soluble and
15	reactive.

17 3.5 DOP and DOS

The relationship between sulfide and iron plays an important role in controlling of the sulfur cycle. As shown in Table 1, both DOP and DOS for the three areas are generally close to each other, with DOP ranging from 0.01 to 0.31 (averages of 0,02 for the Bohai Sea, and 0.04 for both the Yellow Sea and East China Sea) and DOS ranging from 0.02 to 0.33 (averages of 0.04 for the Bohai Sea, 0.05) for the Yellow

1	Sea and 0.04 for the East China Sea. DOS values are slightly higher than the DOP
2	values, which perhaps reflects the reaction of FeS and H_2S to form pyrite (Rozan et al.,
3	2002). The three areas have much lower DOP and DOS in comparison with the
4	averages for estuarine and bay sediments of the Gulf of Mexico (DOP 0.56-0.95 and
5	DOS 0.63-2.66) (Morse et al., 2007), but are consistent with DOP (0.05-0.25) and
6	DOS (0.05-0.39) for Jiaozhou Bay sediment in the Yellow Sea (Zhu et al., 2012). On
7	the basis of the simple comparison above, it is inferred that maybe there was low
8	availability of labile organic matter in the study areas, which has been limiting sulfide
9	production (Zhu et al., 2012), resulting in low values of DOP and DOS. Furthermore,
10	high concentrations of reactive iron (average is approximately 100 μ mol g ⁻¹) and low
11	values of DOP and DOS found in the present study further indicate that pyritization
12	and sulfidization are not being limited by the availability of reactive Fe. Thus in these
13	sediments the supply of mineral-Fe has been amply sufficient to match net sulfide
14	production and thus limit the sulfide toxicity hazard.
15	
16	Table 1
17	
18	4. Conclusions
19	In order to elucidate the geochemical processes controlling the formation and
20	stability of RIS species and its bioavailability in surface sediments, we have examined
21	the relationship between the spatial variations in the values of sedimentary grain size,
22	TOC, TN, TS, AVS, CRS, ES, C/N ratio, C/S ratio, AVS/CRS ratio, DOP and DOS in

1	Bohai Sea, Yellow Sea and East China Sea. In the study area, CRS was the dominant
2	fraction of RIS, indicating the RIS is dominantly in a stable form and thus poses a low
3	threat of toxicity to marine ecosystem. Low values of DOP and DOS indicate that
4	pyritization and sulphidization are not being limited by the availability of reactive Fe.
5	Sediment in these areas restricts the threat from sulfide to benthic organisms. An oil
6	spill in part of the studied area has resulted in enhanced accumulation of AVS in
7	marine sediment, resulting in a higher toxicity risk.
8	
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14	
15	References
16	Bagarinao T., 1992. Sulfide as an environmental factor and toxicant: tolerance and
17	adaptations in aquatic organisms, Aquatic Toxicology 24, 21-62.
18	Berner, R.A., 1970. Sedimentary pyrite formation. American Journal of Science 268,
19	1-23.
20	Berner, R.A., 1982. Burial of organic carbon and pyrite sulphur in the modern ocean:
21	its geochemical and environmental significance. American Journal of Science 282,
22	451-473.

1	Berner, R.A., 1984. Sedimentary pyrite formation: an update. Geochimica et
2	Cosmochimica Acta 48, 605-615.
3	Boesen, C., Postma, D., 1988. Pyrite formation in anoxic environments of the Baltic.
4	American Journal of Science 288, 575-603.
5	Bottrell, S.H., Newton, R.J. 2006. Reconstruction of changes in global sulfur cycling
6	from marine sulfate isotopes. Earth-Science Reviews 75, 59-83.
7	Bottrell S.H., Mortimer R.J.G, Davies I.M., Harvey S.M., Krom M.D., 2009. Sulphur
8	cycling in organic-rich marine sediments from a Scottish fjord. Sedimentology 56,
9	1159-1173.
10	Burton, E.D., Bush, R.T., Johnston, S.G., Sullivan, L.A., Keene, A.F. 2011. Sulfur
11	biogeochemical cycling and novel Fe-S mineralization pathways in a tidally
12	re-flooded wetland. Geochimica et Cosmochimica Acta 75, 3434-3451.
13	Butler, I.B., Rickard, D. 2000. Framboidal pyrite formation via the oxidation of iron
14	(II) monosulfide by hydrogen sulfide. Geochimica et Cosmochimica Acta 64,
15	2665-2672.
16	Canfield, D.E., Thamdrup, B., 1994. The production of ³⁴ S-depleted sulfide during
17	bacterial disproportionation of elemental sulfur. Science 266, 1973-1975.
18	Canfield, D.E., Raiswell, R., Bottrell, S., 1992. The reactivity of sedimentary iron
19	minerals toward sulfide. American Journal of Science 292, 659-683.
20	Canfield, D.E., Kristensen, E., Thamdrup, B., 2005. The Sulfur Cycle, Advances in
21	Marine Biology 48, 313-381.
22	Hsieh Y. P., Shieh Y. N., 1997. Analysis of reduced inorganic sulfur by diffusion

1	methods: improved apparatus and evaluation for sulfur isotopic studies. Chemical
2	Geology, 137, 255-261.
3	Huerta-Dı'az, M.A., Morse, J.W., 1990. A quantitative method for determination of
4	trace metal concentrations in sedimentary pyrite. Marine Chemistry 29, 119-144.
5	Hurtgen, M.T., Lyons, T.W., Ingall, E.D., Cruise, A.M. 1999. Anomalous
6	enrichments of iron monosulfide in euxinic marine sediments and the role of H_2S in
7	iron sulfide transformations: examples from Effingham Inlet, Orca basin, and the
8	Black Sea. American Journal of Science 299, 556-588.
9	Kang, X., Liu, S. and Zhang, G. 2014. Reduced inorganic sulfur in the sediments of
10	the Yellow Sea and East China Sea. Acta Oceanologica Sinica 33,100-108.
11	Lin, S., Huang, KM., Chen, SK., 2000. Organic carbon deposition and its control
12	on iron sulfide formation of the southern East China Sea continental shelf
13	sediments. Continental Shelf Research 20, 619-635.
14	Lin, S., Huang, K. M., Chen, S. K., 2002. Sulfate reduction and iron sulfide mineral
15	formation in the southern East China Sea continental slope sediment. Deep Sea
16	Research Part I: Oceanographic Research Papers 49, 1837-1852.
17	Liu H., Yin B., 2007. Annual cycle of carbon, nitrogen and Phosphorus in the Bohai
18	Sea: A model study, Continental Shelf Research 27, 1399-1407.
19	Liu D., Keesing J.K, He P., Wang Z., Shi Y., Wang Y., 2013. The world's largest
20	macroalgal bloom in the Yellow Sea, China: Formation and implications, Estuarine,
21	Coastal and Shelf Science 129, 2-10.
22	Lv Y., Zhang W., Gao Y., Ning S., Yang B., 2011. Preliminary study on responses of

1	marine nematode community to crude oil contamination in intertidal zone of Bathing
2	Beach, Dalian, Marine Pollution Bulletin, 62, 2700-2706.
3	Morgan, B., Burton, E.D., Rate, A.W., 2012. Iron monosulfide enrichment and the
4	presence of organosulfur in eutrophic estuarine sediments, Chemical Geology
5	296-297, 119-130.
6	Morse, J.W., 1999. Sulfides in sandy sediments: new insights on the reactions
7	responsible for sedimentary pyrite formation. Aquatic Geochemistry 5, 75-85.
8	Morse, J.W., Berner, R.A., 1995. What determines sedimentary C-S ratios. Geochim.
9	Cosmochim. Acta 59, 1073-1077.
10	Morse, J.W., Rickard, D., 2004. Chemical dynamics of sedimentary acid volatile
11	sulfide. Environmental Science & Technology 38, 131A-136A.
12	Morse, J.W., Thomson, H., Finneran, D.W., 2007. Factors controlling sulfide
13	geochemistry in sub-tropical estuarine and bay sediments. Aquatic Geochemistry 13,
14	143-156.
15	Newton R. J., Bottrell S.H., Dean S.P., Hatfield D., Raiswell R., 1995. An evaluation
16	of the use of the chromous chloride reduction method for isotopic analyses of pyrite
17	in rocks and sediment, Chemical Geology 125, 317-320.
18	Phillips B. M., Anderson B. S., Hunt J. W., 1997. Measurement and distribution of
19	interstitial and overlying water ammonia and hydrogen sulfide in sediment, Marine
20	Environmental Research 44, 117-126.
21	Poulton, S.W. 2003. Sulfide oxidation and iron dissolution kinetics during the
22	reaction of dissolved sulfide with ferrihydrite. Chemical Geology 202, 79-94.

1	Qiao S, Shi X, Zhu A, Liu Y., Bi N., Fang X., Yang G., 2010, Distribution and
2	transport of suspended sediments off the Yellow River (Huanghe) mouth and the
3	nearby Bohai Sea. Estuarine, Coastal and Shelf Science 86, 337–344.
4	Rickard, D., Morse, J.W., 2005. Acid volatile sulfide (AVS). Marine Chemistry 97,
5	141-197.
6	Rodrigues S.K., Abessa D.M.S., Machado E.C., 2013. Geochemical and
7	ecotoxicological assessment for estuarine surface sediments from Southern Brazil,
8	Marine Environmental Research 91, 68-79.
9	Rozan, T.F., Taillefert, M., Trouborst, R.E., Glazer, B.T., Ma, S. 2002.
10	Iron-sulfur-phosphorous cycling in the sediments of a shallow coastal
11	bay:implications for sediment nutrient release and benthic macro-algal blooms.
12	Limnology and Oceanography 47, 1346-1354.
13	Schoonen, M.A.A., Barnes, H.L. 1991. Mechanisms of pyrite and marcasite formation
14	from solution: III Hydrothermal processes. Geochim. Cosmochim. Acta 55,
15	3491-3504.
16	Sheng, Y., Fu, G., Chen F., Chen J., 2011.Geochemical characteristics of inorganic
17	sulfur in Shijing River, South China. Journal of Environmental Monitoring 13, 807
18	- 812.
19	Sheng, Y., Sun, Q., Bottrell, S.H., Mortimer, R.J.G., Shi, W., 2013. Anthropogenic
20	impacts on reduced inorganic sulfur and heavy metals in coastal surface sediments,
21	north Yellow Sea, Environmental Earth Sciences 68, 1367-1374.
22	Simpson, S.L., Ward, D., Strom, D., Jolley, D.F., 2012. Oxidation of acid-volatile

1	sulfide in surface sediments increases the release and toxicity of copper to the
2	benthic amphipod Melita plumulosa, Chemosphere 88, 953-961.
3	Sorokin, Y.I., Zakuskina, O.Y., 2012. Acid-labile sulfides in shallow marine bottom
4	sediments: A review of the impact on ecosystems in the Azov Sea, the NE Black
5	Sea shelf and NW Adriatic lagoons, Estuarine, Coastal and Shelf Science 98,
6	42-48.
7	Tarpgaard, I.H., Røy, H., Jørgensen, B.B., 2011. Concurrent low- and high-affinity
8	sulfate reduction kinetics in marine sediment, Geochimica et Cosmochimica Acta,
9	75, 2997-3010.
10	Xu, K., Milliman, J.D., Li, A., Liu, J.P., Kao, SJ., Wan, S., 2009. Yangtze- and
11	Taiwan-derived sediments on the inner shelf of East China Sea. Continental Shelf
12	Research 29, 2240–2256.
13	Yin, H., Fan, C., Ding, S., Zhang, L. and Zhong, J., 2008. Geochemistry of Iron,
14	Sulfur and Related Heavy Metals in Metal-Polluted Taihu Lake Sediments,
15	Pedosphere 18, 564-573.
16	Zhou, J., Wu, Y., Kang, Q., Zhang, J., 2007. Spatial variations of carbon, nitrogen,
17	phosphorous and sulfur in the salt marsh sediments of the Yangtze Estuary in China,
18	Estuarine, Coastal and Shelf Science 71, 47-59.
19	Zhu, M., Liu, J., Yang, G., Li, T., Yang, R., 2012. Reactive iron and its buffering
20	capacity towards dissolved sulfide in sediments of Jiaozhou Bay, China, Marine
21	Environmental Research 80, 46-55.
22	Zhu, M., Shi X., Yang G., Hao X., 2013. Formation and burial of pyrite and organic
	21

sulfur in mud sediments of the East China Sea inner shelf: Constraints from
 solid-phase sulfur speciation and stable sulfur isotope, *Continental Shelf Research* 54, 24-36.

Figure captions 1

2

7	B30, B31 and B32 were illustrated with the different scales)
6	Fig.4 Concentrations of different fractions of RIS in surface sediments (Note: si
5	Fig. 3 Concentrations of TOC, TN and TS in surface sediment in different locations
4	Fig. 2 Grain size distribution of surface sediments in different sampling sites
3	Fig. 1 Location of studying areas and sampling sites

8

(Note: sites

Yellow Sea	C/N	C/S	AVS/CRS	DOP	DOS	Fe _R	Fe _{py}	Bohai Sea	C/N	C/S	AVS/CRS	DOP	DOS	Fe _R	Fe _{py}
B01	7.84	6.67	0.08	0.04	0.04	147.43	5.70	B40	15.07	10.69	0.22	0.01	0.02	66.97	1.21
B02	11.92	14.70	0.23	0.03	0.05	113.41	3.85	B41	16.01	9.46	0.90	0.03	0.06	76.03	1.11
B03	17.22	11.81	0.94	0.02	0.07	65.53	1.12	B43	17.68	10.82	0.77	0.02	0.05	100.66	2.80
B04	16.43	13.29	0.12	0.01	0.01	110.69	0.77	B44	19.49	11.99	0.74	0.02	0.05	92.21	2.08
B05	16.09	17.28	0.17	0.03	0.04	71.51	2.04	B45	23.36	16.13	0.43	0.01	0.03	91.21	1.95
B07	12.99	8.74	0.07	0.04	0.05	85.19	3.72	B46	18.79	15.11	0.40	0.03	0.05	110.01	1.64
B09	10.00	11.28	0.09	0.03	0.04	65.35	2.17	B47	16.95	12.58	1.22	0.01	0.03	110.69	3.71
B13	12.52	3.50	0.05	0.09	0.10	46.95	4.39	B48	18.81	15.97	0.43	0.01	0.03	94.68	1.27
B15	8.34	1.31	0.15	0.02	0.03	22.56	0.39	B49	13.68	9.35	0.46	0.01	0.03	43.94	0.35
B21	9.71	5.35	0.14	0.03	0.05	50.42	1.73	B50	13.90	12.94	1.19	0.01	0.10	83.22	1.13
B23	11.58	5.45	0.06	0.04	0.05	63.96	3.00	B51	13.27	7.89	0.25	0.01	0.02	39.03	0.31
B24	10.40	6.81	0.16	0.03	0.04	100.27	3.19	B52	10.07	11.13	0.15	0.01	0.01	117.96	1.67
B25	9.60	9.07	0.08	0.06	0.07	84.73	5.43	B53	23.58	10.76	0.15	0.02	0.03	124.98	1.48
B26	18.50	11.33	0.53	0.01	0.03	69.19	1.03	B54	15.51	6.17	0.20	0.03	0.04	66.43	1.20
B28	24.53	18.66	0.33	0.01	0.02	56.19	0.43	B55	12.73	6.42	0.16	0.01	0.04	91.50	3.10
B30	8.16	2.60	0.01	0.27	0.28	77.35	28.79	B56	15.11	6.99	0.19	0.02	0.02	65.25	0.97
B31	41.32	12.47	0.02	0.31	0.33	39.32	17.74	B59	14.94	9.62	0.49	0.02	0.04	83.94	1.65
B32	15.81	4.82	0.04	0.28	0.32	76.56	30.24	B60	14.90	12.83	0.14	0.04	0.04	78.57	1.65
B33	20.87	10.37	0.19	0.03	0.07	28.29	0.98	B61	11.97	6.64	0.26	0.03	0.04	113.02	4.93
B34	16.08	9.35	0.40	0.01	0.02	40.57	0.54	B62	10.48	10.93	0.16	0.02	0.03	138.62	3.62
B37	14.10	12.85	0.28	0.02	0.05	53.68	1.13	B63	13.32	10.62	0.18	0.02	0.02	124.58	1.91
B38	15.29	13.56	0.39	0.02	0.03	70.55	1.29	B64	18.40	12.39	0.77	0.02	0.03	114.49	2.04
H01	24.94	24.28	0.64	0.01	0.01	102.20	0.79	B66	29.69	37.48	0.83	0.03	0.05	71.33	1.19
H02	22.77	24.22	0.16	0.01	0.01	74.91	0.62	B67	27.41	13.18	0.14	0.01	0.02	88.67	2.65

Table 1 Calculation results of different measured geochemical parameters and different iron concentrations (µmol g⁻¹).

H03	19.12	15.18	0.42	0.00	0.01	71.08	0.17	B68	34.54	12.45	0.20	0.02	0.02	66.03	0.69
H04	11.89	9.41	0.51	0.01	0.02	110.87	1.55	B69	20.17	12.28	0.93	0.02	0.02	94.15	1.71
H05	8.87	7.28	0.06	0.02	0.03	165.16	4.22	B 70	21.00	15.01	1.20	0.04	0.04	91.89	1.60
H06	8.32	6.14	1.09	0.02	0.03	163.58	3.16	YD1	48.41	30.10	0.36	0.01	0.02	77.82	0.68
H07	7.71	6.92	0.05	0.02	0.02	172.32	3.55	Mean	18.90	12.78	0.48	0.02	0.04	89.92	1.80
H08	8.35	7.45	0.18	0.02	0.03	185.00	3.82	East China Sea							
H09	8.25	6.32	0.08	0.04	0.04	159.28	6.35	H42	16.81	6.52	0.08	0.04	0.05	127.45	5.76
H10	8.57	11.20	0.10	0.03	0.03	150.15	4.57	E01	16.73	13.32	0.11	0.03	0.04	171.85	4.92
H11	9.58	5.87	0.02	0.04	0.04	179.19	7.25	E02	18.69	16.97	0.09	0.04	0.05	111.37	4.74
H12	9.69	7.40	0.12	0.02	0.03	164.26	3.24	E03	17.96	6.50	0.04	0.03	0.03	148.40	4.47
H14	10.80	10.14	0.46	0.01	0.03	91.82	1.04	E04	20.75	8.45	0.03	0.03	0.03	110.37	3.67
H16	16.18	7.27	0.96	0.01	0.02	52.46	0.59	E05	15.64	10.20	0.04	0.04	0.04	111.37	4.76
H43	9.26	9.02	0.11	0.03	0.04	181.77	5.49	Mean	17.76	10.33	0.07	0.04	0.04	130.14	4.72
H44	9.10	10.15	0.15	0.02	0.03	178.48	4.47								
HF1	12.48	11.32	0.13	0.01	0.02	50.35	0.54								
HF3	8.34	8.38	0.05	0.03	0.04	138.33	4.48								
H29	12.53	12.26	0.08	0.02	0.03	140.30	3.29								
H31	14.47	7.65	0.18	0.03	0.04	134.50	3.53								
H32	17.42	13.03	0.08	0.02	0.03	147.61	3.20								
H33	24.68	8.46	0.07	0.02	0.04	108.76	2.76								
H34	20.65	8.10	0.07	0.03	0.03	140.27	3.69								
H35	40.87	21.72	0.07	0.02	0.02	105.43	1.64								
H36	33.38	16.84	0.17	0.02	0.03	98.98	2.00								
H38	20.68	13.88	0.08	0.02	0.02	139.05	2.91								
Mean	15.17	10.44	0.22	0.04	0.05	103.04	4.14								



Fig. 1



Fig. 2



Fig. 3



Fig.4