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### **Title**

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**Title: Micronutrients intake is associated with improved sperm DNA quality in older men**

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**Capsule:** Older men with higher dietary and supplement intake of certain micronutrients including vitamin C, vitamin E, folate, and zinc, but not  $\beta$ -carotene, may produce sperm with less DNA damage.

**Abstract (249 words)**

**OBJECTIVE:** To investigate whether lifestyle factors such as increased dietary intake of micronutrients reduce the risks of sperm DNA damage, and whether older men benefit more than younger men.

**DESIGN:** Non-clinical group of 22-80-year-old non-smoking men (n~80) who reported no fertility problems.

**METHODS:** Sociodemographics, occupational exposures, medical and reproductive histories, and lifestyle habits were determined by questionnaire. The average daily dietary and supplement intake of micronutrients (vitamin C, vitamin E,  $\beta$ -carotene, zinc, and folate) was determined using the 100-item Modified Block Food Frequency Questionnaire (FFQ). Associations between estimated micronutrient intake and sperm DNA damage, as well as their interactions with male age, were then assessed.

**MAIN OUTCOME MEASURES:** sperm DNA damage measured by alkaline and neutral DNA electrophoresis, i.e., sperm Comet.

**RESULTS:** Men with highest intake of vitamin C had ~16% less sperm DNA damage (alkaline sperm Comet) than men with lowest intake ( $p < 0.05$ ), with similar findings for vitamin E, folate, and zinc (but not  $\beta$ -carotene). Older men ( $>44y$ ) with highest vitamin C intake had ~20% less sperm DNA damage compared with older men with lowest intake ( $p < 0.05$ ), with similar findings for vitamin E and zinc. However, younger men ( $<44y$ ) did not benefit from higher intakes of the micronutrients surveyed.

**CONCLUSIONS:** Men with higher dietary and supplement intake of certain micronutrients may produce sperm with less DNA damage, especially among older men. This raises the broader question of how lifestyle factors including higher intakes of antioxidants and micronutrients might protect somatic as well as germ cells against age-associated genomic damage.

**Key words:** human male aging; micronutrients and antioxidant; sperm alkaline and neutral Comet assay; human sperm DNA damage; vitamin C, vitamin E,  $\beta$ -carotene, zinc, and folate

## Introduction

The increasing trend for men over 35 years of age to have children raises public health concern about age-associated risks for abnormal pregnancies and birth defects. Male fecundity is known to decline with age, although spermatogenesis continues well into male senescence and some men of substantially advanced ages have fathered healthy children (1, 2). Nevertheless, men's age has been associated with reduced semen quality (3), increased frequencies of sperm with structural aberrations (4), increased sperm DNA fragmentation and increased frequencies of sperm carrying achondroplasia gene mutations (5). Thus, normally aging males are predicted to experience higher frequencies of chromosomally defective pregnancies and higher proportions of offspring carrying heritable genetic defects (5). In addition, male aging has been associated with sperm DNA strand damage (6), but no studies have examined whether lifestyle and diet can be protective against detrimental effects of aging on the sperm genome.

Dietary supplementations with antioxidants and increased intake of fruits or vegetables rich in antioxidants have been shown to decrease the amount of oxidative DNA damage *in vivo* (7, 8) and to reduce DNA breaks (9) as well as DNA oxidation (7) in human lymphocytes. *In vitro* evidence shows that vitamin C (ascorbic acid), a scavenger of reactive oxygen species, can reduce oxidative stress in sperm (10). Our laboratory reported that antioxidant intake was associated with better semen quality, in particular, sperm motility (11), and that men with higher folate intake were less likely to produce aneuploid sperm for chromosomes X, Y and 21 (12). However, no association was observed between antioxidant intake and DNA fragmentation (13). Whether antioxidant intake and general dietary changes can enhance male fertility and improve



reproductive outcomes is a research topic in need of better understanding of underlying cellular and molecular mechanisms (14-16).

Our current study utilizes a population of healthy non-smokers in which we previously reported age-associated increases of sperm DNA damage using the sperm Comet assay (6) to address two questions: (a) is micronutrient intake differentially associated with either of the two subtypes of sperm DNA damage measured by the sperm Comet assay (alkaline and neutral versions; (17))? and (b) does micronutrient intake modify the age-related increases we previously observed? The alkaline version of the sperm Comet assay is thought to detect double strand breaks (DSB), single strand breaks (SSB) and alkali-labile sites (ALS) whereas the neutral version is thought to predominantly detect DSB and to a lesser extent SSB (17).

## **Material and Methods**

### *Study population.*

The study population consisted of ~80 healthy male volunteers, aged 22-80, who were recruited for the California Age and Genetic Effects on Sperm (AGES) Study. Participants were recruited from among employees and retirees of a national laboratory in California, which provides health care plans for its employees. Recruitment was conducted through the University of California, Berkeley, and the sperm analysis was performed in a research setting at the national laboratory. The California AGES Study was approved by the Institutional Review Board of each participating institution, and all volunteers gave their written consent to participate. Preliminary screenings excluded men who had current fertility or reproductive problems; had smoked cigarettes in the last six months; had a vasectomy or a history of an undescended testicle or prostate cancer; had received chemotherapy or radiation treatments for cancer; or, had a previous semen analysis with zero sperm count. If men at screening had a fever over 38.5 °C in the last three months, they were enrolled three months from the date of the fever. At least 15 men were enrolled for each age decade between 20 and 70 years; additional men above age 70 were also enrolled. Of the original set of men in the California AGES Study, one subject missing dietary data was excluded. Additional subjects were excluded if they did not have adequate remaining semen aliquots for Comet analyses, resulting in a total of 79 men for the analysis of neutral sperm comet assay and 78 for the alkaline sperm comet assay. Sixty-two percent of the men had fathered a pregnancy. Men who fathered pregnancy were older than men who had not (48.8 vs. 35.3 years,  $p < 0.001$ ).

### *Questionnaire.*

We mailed to each eligible participant a study questionnaire, semen collection instructions, a sterile container, and protective thermos (11, 13). The self-administered questionnaire asked about sociodemographic characteristics (age, race, and education), occupation, possible occupational exposures, medical and reproductive history, and lifestyle habits. We also sent a 100-item Modified Block Food Frequency Questionnaire (FFQ) (18) to estimate average daily dietary and supplemental vitamin intake. The FFQ assessed the frequency and serving size of major food types over the previous year, including specific supplement intake information for: multivitamins; antioxidant combination vitamins; calcium; iron; zinc; and selenium. FFQ responses were converted into average daily nutrient intake estimates using a standardized reference nutrient database ([www.nutritionquest.com](http://www.nutritionquest.com)). Participants completed the FFQ within one week of producing the semen sample. Study staff reviewed the FFQs for reasonable levels of calories and number of foods per day. Questionnaires were reviewed with participants over the phone for completion and accuracy.

### *Semen Collection and Sperm Comet Assay.*

Men provided a semen sample by masturbation into a sterile collection container after abstinence of 2-7 days, and delivered it within an insulated thermos at room temperature to a drop box at the laboratory, noting the actual duration of abstinence. Samples were delivered within two hours of collection and immediately analyzed for conventional semen quality and computer assisted semen analysis (CASA) (19).

Remaining aliquots of semen were stored at  $-80^{\circ}$  C. Frozen samples were shipped on dry ice to the University of Bradford, United Kingdom and kept at  $-80^{\circ}$  C until analysis. The sperm Comet data used in this analyses were previously published (6), and were based on the prior procedures described by Anderson et al. for the alkaline assay (20) and by Duty et al. for the neutral assay (21). Slides were examined at 400X magnification on a fluorescent microscope. Fifty cells were scored from each replicate slide (100 cells total). A computerized image analysis system (Comet 3.0, Kinetic Imaging) was used to measure the percent tail DNA and other parameters including tail extent moment and olive tail moment. These different measurements were highly correlated in our dataset and in the literature. We calculated the percent DNA in the comet tail because it is a clear indication of sperm comets, linearly related to the DNA break frequency over a wide range of levels of damage, the most commonly reported outcome (22), and considered a reliable measurement (23) for human sperm samples.

#### *Statistical analysis.*

The main independent variables were estimated daily intake of vitamin C, vitamin E, beta-carotene, folate and zinc as well as a composite of antioxidant intake. Intake was based on the combination of reported diet and supplements use and was divided into: low (less than 25th percentile), moderate (25th to 75th percentile), and high (greater than 75th percentile). Quartiles were created using all AGES study participants with complete FFQ data to ensure that an individual remained in the same category for all AGES study analyses (11-13). The antioxidant composite variable was created *a priori* to obtain a combined measure of dietary and supplementary antioxidant intake from vitamins C, E

and beta-carotene, as described previously (11). Low antioxidant composite intake was defined as low intake for at least two of the antioxidants; high antioxidant composite intake was defined as high intake for at least two of the antioxidants; and moderate intake was defined as all other combinations of intake.

All analyses were performed using Stata 8.0 (24). Separate multivariable linear regression models were constructed to examine the relationship of intake of each antioxidant with sperm DNA damage (i.e., %tail DNA) for the neutral and alkaline Comet assays. For each nutrient, the three intake groups were included as indicator variables into statistical models, with low intake as baseline. The following variables and classes of variables were considered potential covariates: age; time from sample collection to sample processing; duration of sexual abstinence before semen collection; season of sample collection; exposure to occupational chemicals and radiation; history of working with radioisotopes; history of tobacco use; alcohol and caffeine intake; prescription and non-prescription medication use; history of chronic disease such as high blood pressure, heart problems, or diabetes; history of genitourinary disease including urinary tract or other genitourinary infection, sexually transmitted disease, varicocele, or past history of infertility; fatherhood history; body mass index; and ethnicity. Covariates were first examined in univariate analyses with sperm DNA damage using analysis of variance (ANOVA) and *t*-tests and were included in the full models if  $p < 0.2$ . Covariates were retained in the final models if they changed the nutrient regression coefficient by at least 10% or had a  $p$ -value  $< 0.10$ . We checked regression assumptions with residual vs. fitted plots, and quantile-quantile plots. Final regression results are expressed as adjusted means and standard errors (SE) predicted from the models with covariates set at their

mean values. To obtain p-values for trend, in the final models we replaced the nutrient indicator variables with ordered variables (i.e., 1=low intake, 2=moderate, 3=high). All p-values presented are two-sided.

To test whether higher micronutrient intake affected the positive relationship of age and alkaline sperm DNA damage reported previously (6), we created interaction terms for age (dichotomized at the median age of 44) and nutrient levels (dichotomized at the median intake for each nutrient) and assessed the associations with sperm DNA damage from the alkaline Comet assay. Any model with an interaction term  $<0.2$  was considered statistically significant and was re-analyzed using dummy variables for the different intake/age groups using old age/low intake as the referent group. Results are expressed as adjusted means and 95% confidence intervals (CI), (Table 3).

Four men with high sperm DNA damage (i.e., alkaline % tail DNA values  $>2.5$  SD from mean) were evaluated as potential outliers (6). Three of the four men were  $>65$  years of age, all four had fathered children and none had a history of fertility problems. All alkaline comet models were analyzed with and without these four men and results of all models are presented.

## Results

### *Characteristics of study population*

Men were on average 46.4 years old (range 22 to 80 years), generally healthy, and had not smoked cigarettes during the previous six months. The magnitude of sperm DNA damage was  $42.2 \pm 8.9\%$  (% tail DNA, mean  $\pm$  SD) and  $35.2 \pm 7.9\%$  under alkaline conditions and neutral conditions, respectively, with no significant correlation between the results of the two analyses (Pearson correlation,  $p > 0.1$ ), (6). As summarized in Table S1 (see Supplemental Material), we found that higher DNA damage in the alkaline assay was associated with age ( $p < 0.05$ ) and with history of kidney, bladder and urinary tract infections ( $p < 0.05$ ). Under neutral conditions, higher sperm DNA damage was associated with higher body mass index ( $p < 0.05$ ).

### *Micronutrient intake and sperm DNA damage*

The median daily total intake (diet + supplements) was 162 mg for vitamin C, 23.7 mg  $\alpha$ -tocopherol equivalents ( $\alpha$ TE) for vitamin E, 2,586  $\mu$ g for  $\beta$ -carotene, 475  $\mu$ g for folate, and 12.3 mg for zinc (Table 1). The percent of men consuming less than the recommended dietary allowance (RDA) for each micronutrient ranged from 22% (vitamin C) to 47% (zinc) for total intake (dietary intake and supplements) and from 39% (vitamin C) to 81% (vitamin E) for dietary intake only. All further analyses were performed by using the measure of combined diet and supplement intakes.

We found no evidence of significant associations between any of the measures of micronutrient intake and sperm DNA damage as measured by the neutral version of the sperm Comet assay. Thus, the further analyses were conducted only between

micronutrient intact and sperm DNA damage as measured by the alkaline version of the sperm Comet assay.

Vitamin C, E, beta-carotene. As shown in Table 2, after adjusting for age and other covariates (see methods), men in the highest quartile of daily vitamin C intake (459-3370 mg per day) had 16% less sperm DNA damage in the alkaline Comet assay ( $p < 0.01$ ) than men in the lowest quartile (26-99 mg per day) with a significant test for trend across the three intake categories ( $p < 0.01$ ). For vitamin E, there was a similar pattern as men with the highest intake had lower sperm DNA damage than men with the lowest intake, although the relationship was non-significant ( $p = 0.11$ , test for trend  $p = 0.12$ ). In addition, the antioxidant composite measure (Vitamin C, Vitamin E, Beta-Carotene) showed a decrease in sperm DNA damage between the highest intake group and the lowest intake group (39.1 vs. 45.9,  $p = 0.03$ ) with a significant trend ( $p = 0.02$ ). There was no significant association between beta-carotene and sperm DNA damage.

When we excluded the four outliers with high values of sperm DNA damage (i.e., alkaline % tail DNA  $> 60\%$ ), we still observed a significant inverse relationship between sperm DNA damage and nutrient intake for vitamin C ( $p < 0.01$ ) and the antioxidant composite variable ( $p = 0.05$ ), as well as the borderline-significant relationship with vitamin E (test for trend,  $p = 0.10$ ). Excluding the outliers did not change the relationship of sperm DNA damage with beta-carotene.

Folate. Men in the highest quartile of high folate intake (722-1,150 ug per day) had lower levels of sperm DNA damage compared to men in the lowest quartile (114-333 ug



per day), albeit non-significantly (39.3 vs. 43.5,  $p=0.13$ ). When the four outliers were excluded from the analysis, this relationship strengthened and became statistically significant (38.0 vs. 42.1,  $p=0.05$ ).

*Zinc.* As shown in Table 2, after adjusting for covariates, men in the highest quartile of daily zinc intake (22-74 mg per day) had less sperm DNA damage in the alkaline Comet assay ( $p = 0.07$ ) than men in the lowest quartile (5-8 mg per day) (test for trend,  $p=0.07$ ). The relationship was strengthened slightly when the outliers were excluded ( $p=0.04$  for both high vs. low comparison and test for trend).

#### *Interaction with age.*

In models where nutrient intake was related to sperm DNA damage, age also remained significantly related, suggesting that nutrient intake did not eliminate the age relationship with sperm DNA damage reported previously (6). In further examination of the relationship between age and intake on sperm DNA damage, there was a consistent finding for vitamin C, vitamin E, zinc, and folate (but not for betacarotene) that the older men with the lower intake (intake < median level for the micronutrient) had the highest sperm DNA damage compared to the other groups, including older men with higher intake (Table 3); this association was not significant for folate. Furthermore, older men with higher intake had sperm DNA damage comparable to the younger men. Figure 1 shows the relation between age and sperm DNA damage for vitamin C, E, and zinc.

The difference in sperm DNA damage for older men with high zinc intake compared to older men with low zinc intake remained statistically significant even after

excluding the four outliers. However, sperm DNA damage for older men with high vitamin C and vitamin E intake was now non-significantly lower than older men with low intake ( $p=0.11$  and  $0.12$ , respectively) but still similar to values for younger men with high intake even when excluding the four outliers ( $p=0.04$ ).

## Discussion

We previously reported significant associations between vitamin C, vitamin E,  $\beta$ -carotene intake and semen quality (11). We also reported that older men had more sperm DNA damage using the comet assay (6). In the present study, we report that (1) *men with the higher total intake of vitamin C, vitamin E, folate and zinc from diet and supplements had significantly less sperm DNA damage (alkaline Comet) than men with lower intake and (2) older men who consumed higher levels of certain micronutrients (vitamin C, vitamin E, and zinc) had similar levels of sperm DNA damage than young men.* These new findings suggest that for men who are at increased risk of DNA strand damage due to advancing age, a diet that consists of high levels of antioxidants and micronutrients may decrease the risk of producing sperm with DNA damage.

Our findings that high antioxidant intake was associated with lower amounts of DNA breaks may be explained by changes in oxidative stress in the male reproductive tract (25, 26). Anderson et al. showed that vitamin C can be an antioxidant in the sperm Comet assay (27). Vitamin C, a scavenger of reactive oxygen species (ROS) (28), is secreted from the seminal vesicles during ejaculation, and is the major antioxidant present in seminal plasma of fertile males, contributing to up to 65% of the total antioxidant capacity (29). In a prior study, we showed that dietary intake of Vitamin C can protect human sperm against endogenous oxidative DNA damage (30). Hughes and colleagues reported that *in vitro* treatment of sperm with alpha-tocopherol, an antioxidant, reduced the magnitude of DNA damage as measured by the Comet assay (31).

Folate, another antioxidant, also appears to affect genetic quality of sperm in older men. Folate is an essential B-vitamin that occurs naturally in a wide variety of foods,

such as broccoli, cabbage, cauliflower, fruit and nuts. Folate metabolism is critical for proper cell function (32, 33). Its main roles are to provide carbon groups for purine and pyrimidine synthesis and to create the vital methyl donors methionine and S-adenosylmethionine (34). A low thymidine/uracil ratio caused by folate deficiency results in uracil being incorporated into DNA during synthesis. The repair of two nearby opposing lesions can cause DNA double strand breaks (35). It was shown that dietary deficiency in folate increased DNA strand breaks in the liver of rats (36). Folate deficiency has been also shown to increase chromosomal instability and aneuploidy (32, 33) in human lymphocytes. Physiological levels of folate deficiency can cause DNA damage in human lymphocytes, by up-regulation of several excision repair genes and down-regulation of many folate metabolism genes (35). There is an interaction between folic acid deficiency and oxidative stress on DNA damage because the simultaneous excision of an oxidized base and uracil on opposite DNA strands within 12 bases of each other could result in the formation of double-strand breaks (37).

Lopes et al. (38) showed that *in vitro* exposure of human sperm to ROS increased DNA fragmentation as measured by the TUNEL assay, while *in vitro* treatment with antioxidants (including cysteine and glutathione) reduced DNA fragmentation. Oxidative stress can induce oxidative damage in the sperm plasma membrane and DNA damage in mitochondrial and nuclear genomes (39). Kodama and colleagues reported an association between the level of endogenous oxidative DNA damage in spermatozoa and male infertility linked to sperm DNA damage (40). The association of oxidative damage and infertility is supported by our previous findings that antioxidant intake is associated with better semen quality, in particular, motility (11). Mechanistically, antioxidants may be

important for protecting human sperm from peroxidative damage that may result from the high amount of poly-unsaturated fatty acids in sperm (14).

Our current study noted an association between zinc intake and sperm DNA damage as measured by the alkaline sperm Comet assay. Ten percent of the U.S. population consumes less than half the recommended daily allowance for zinc, especially those who eat little meat and/or consume high phytate-containing foods (41). Insufficient zinc intake can impair antioxidant defences and compromise DNA repair, making the cell susceptible to oxidative damage (42). Zinc deficiency can also lead to increased oxidative damage to testicular DNA (43). There is also evidence from rats that zinc deficiency was associated with increased sperm DNA damage (44). Yang and colleagues used the sperm Comet assay to show that higher zinc concentrations can prevent human sperm from damage by sodium nitroprusside, a blood pressure reducing drug (45). Together, these data suggest that zinc deficiency can compromise DNA integrity via oxidative stress, but the mechanisms by which zinc deficiency induces oxidative stress are still unclear. One possibility is that zinc deficiency may impair mitochondrial function, causing excess free radical species to be leaked through the electron transport chain (43).

The protective effects of antioxidants and micronutrient intake were seen in the alkaline version of the sperm Comet assay but not the neutral version, using the same semen specimens. Oxidative stress which increases with age primarily causes SSB and ALS, but only very few DSB. The alkaline Comet assay detects DSB, SSB and ALS whereas the neutral version predominantly detects DSB and to a **much** lesser extent SSB (see review by Baumgartner A., et al 2009). This suggests that antioxidant and micronutrient intake protects specifically against SSB and ALS in sperm which are

increased in aging males (as measured by the alkaline assay), but show no detectable protection for DSBs in sperm which are not increased in aging males (Table 1).

Our biomarker study design had some limitations. First, the AGES study population was composed of healthy non-smoking volunteers without major reproductive problems. Therefore, our findings may not be generalizable to clinic-based groups or infertile populations. Second, our assessment of micronutrient intake was derived from the Modified Block Food Frequency Questionnaire (FFQ). The advantage of the FFQ is that it can ascertain usual intake during the entire period of spermatogenesis (13). However, the FFQ only provides estimates of intake and does not measure longitudinal, daily dietary and supplemental vitamin intake.. Therefore, we cannot assess variation in intake for the separate phases of sperm development. Also, the FFQ was not specifically validated in our population, although it was validated against dietary records in several prior studies (46-48). Third, our cross-sectional design could not determine whether micronutrient intake and sperm DNA damage were causally related; a future randomized clinical trial might address this question. Lastly, because of the correlation among the intake of different micronutrients, we could not definitively determine whether our results derived specifically from a high quality diet, from one nutrient or nutrients in-particular, or from one or several associated life-style factors. . Recently, a position report on sperm DNA organization, protection and vulnerability also highlighted the importance of micronutrients such as zinc for maintaining sperm chromatin integrity (49). Given the potential importance for older men, our findings merit further scrutiny in future dietary studies, including intervention trials.

Our findings suggest that men who consume higher amounts of antioxidants (vitamin C, vitamin E, folate) and the micronutrient zinc will, on average, produce sperm carrying less DNA damage, and that this protective effect is most pronounced among older men. The reproductive and heritable implications for individual older men who take higher amounts has not yet been investigated and it remains to be shown whether these small average changes will enhance reproductive capacity of individuals. Our sperm findings point to a need for future studies to determine whether increased antioxidant intake in older fathers will improve fertility, reduce risks of genetically defective pregnancies, and result in healthier children. Our findings also raise the broader question of whether lifestyle factors including higher intakes of antioxidants and micronutrients can protect germ cells as well as somatic cells against age-related DNA damage.

## References

1. Kidd SA, Eskenazi B, Wyrobek AJ. Effects of male age on semen quality and fertility: a review of the literature. *Fertility and Sterility* 2001;75:237-48.
2. Slotter E, Nath J, Eskenazi B, Wyrobek AJ. Effects of male age on the frequencies of germinal and heritable chromosomal abnormalities in humans and rodents. *Fertil Steril* 2004;81:925-43.
3. Eskenazi B, Wyrobek AJ, Slotter E, Kidd SA, Moore L, Young S *et al.* The association of age and semen quality in healthy men. *Hum Reprod* 2003;18:447-54.
4. Slotter ED, Marchetti F, Eskenazi B, Weldon RH, Nath J, Cabrerros D *et al.* Frequency of human sperm carrying structural aberrations of chromosome 1 increases with advancing age. *Fertil Steril* 2007;87:1077-86.
5. Wyrobek AJ, Eskenazi B, Young S, Arnheim N, Tiemann-Boege I, Jabs EW *et al.* Advancing age has differential effects on DNA damage, chromatin integrity, gene mutations, and aneuploidies in sperm. *Proc Natl Acad Sci U S A* 2006;103:9601-6.
6. Schmid TE, Eskenazi B, Baumgartner A, Marchetti F, Young S, Weldon R *et al.* The effects of male age on sperm DNA damage in healthy non-smokers. *Hum Reprod* 2007;22:180-7.
7. Duthie SJ, Ma A, Ross MA, Collins AR. Antioxidant supplementation decreases oxidative DNA damage in human lymphocytes. *Cancer Res* 1996;56:1291-5.
8. Collins BH, Horska A, Hotten PM, Riddoch C, Collins AR. Kiwifruit protects against oxidative DNA damage in human cells and in vitro. *Nutr Cancer* 2001;39:148-53.
9. Green MH, Lowe JE, Waugh AP, Aldridge KE, Cole J, Arlett CF. Effect of diet and vitamin C on DNA strand breakage in freshly-isolated human white blood cells. *Mutat Res* 1994;316:91-102.
10. Anderson D, Schmid TE, Baumgartner A, Cemeli-Carratala E, Brinkworth MH, Wood JM. Oestrogenic compounds and oxidative stress (in human sperm and lymphocytes in the Comet assay). *Mutat Res* 2003;544:173-8.
11. Eskenazi B, Kidd SA, Marks AR, Slotter E, Block G, Wyrobek AJ. Antioxidant intake is associated with semen quality in healthy men. *Hum Reprod* 2005;20:1006-12.
12. Young SS, Eskenazi B, Marchetti FM, Block G, Wyrobek AJ. The association of folate, zinc and antioxidant intake with sperm aneuploidy in healthy non-smoking men. *Hum Reprod* 2008;23:1014-22.
13. Silver EW, Eskenazi B, Evenson DP, Block G, Young S, Wyrobek AJ. Effect of antioxidant intake on sperm chromatin stability in healthy nonsmoking men. *J Androl* 2005;26:550-6.
14. Wong WY, Thomas CM, Merkus JM, Zielhuis GA, Steegers-Theunissen RP. Male factor subfertility: possible causes and the impact of nutritional factors. *Fertil Steril* 2000;73:435-42.
15. Agarwal A, Nallella KP, Allamaneni SS, Said TM. Role of antioxidants in treatment of male infertility: an overview of the literature. *Reprod Biomed Online* 2004;8:616-27.
16. Sikka SC. Role of oxidative stress and antioxidants in andrology and assisted reproductive technology. *J Androl* 2004;25:5-18.
17. Baumgartner A, Cemeli C, Anderson D. The Sperm Comet Assay - Assessing Genotoxins in Male Germ Cells. In: Dhawan A, Anderson D, eds. *The Comet assay in Toxicology*. Cambridge, UK: Royal Society Chemistry, 2009:331-69.



18. Huang MH, Schocken M, Block G, Sowers M, Gold E, Sternfeld B *et al.* Variation in nutrient intakes by ethnicity: results from the Study of Women's Health Across the Nation (SWAN). *Menopause* 2002;9:309-19.
19. Slotter E, Schmid TE, Marchetti F, Eskenazi B, Nath J, Wyrobek AJ. Quantitative effects of male age on sperm motion. *Hum Reprod* 2006;21:2868-75.
20. Anderson D, Basaran N, Dobrzynska MM, Basaran AA, Yu TW. Modulating effects of flavonoids on food mutagens in human blood and sperm samples in the comet assay. *Teratog Carcinog Mutagen* 1997;17:45-58.
21. Duty SM, Singh NP, Silva MJ, Barr DB, Brock JW, Ryan L *et al.* The relationship between environmental exposures to phthalates and DNA damage in human sperm using the neutral comet assay. *Environ Health Perspect* 2003;111:1164-9.
22. Hartmann A, Agurell E, Beevers C, Brendler-Schwaab S, Burlinson B, Clay P *et al.* Recommendations for conducting the in vivo alkaline Comet assay. 4th International Comet Assay Workshop. *Mutagenesis* 2003;18:45-51.
23. Anderson D, Dobrzynska MM, Basaran N, Basaran A, Yu TW. Flavonoids modulate comet assay responses to food mutagens in human lymphocytes and sperm. *Mutat Res* 1998;402:269-77.
24. StataCorp. Intercooled Stata 8.2 for Windows. In. College Station, TX: Stata Corporation, 2004.
25. Barnes CJ, Hardman WE, Maze GL, Lee M, Cameron IL. Age-dependent sensitization to oxidative stress by dietary fatty acids. *Aging (Milano)* 1998;10:455-62.
26. Barroso G, Morshedi M, Oehninger S. Analysis of DNA fragmentation, plasma membrane translocation of phosphatidylserine and oxidative stress in human spermatozoa. *Hum Reprod* 2000;15:1338-44.
27. Anderson D. Factors that contribute to biomarker responses in humans including a study in individuals taking Vitamin C supplementation. *Mutat Res* 2001;480-481:337-47.
28. Sies H, Stahl W, Sundquist AR. Antioxidant functions of vitamins. Vitamins E and C, beta-carotene, and other carotenoids. *Ann N Y Acad Sci* 1992;669:7-20.
29. Lewis SE, Boyle PM, McKinney KA, Young IS, Thompson W. Total antioxidant capacity of seminal plasma is different in fertile and infertile men. *Fertil Steril* 1995;64:868-70.
30. Fraga CG, Motchnik PA, Shigenaga MK, Helbock HJ, Jacob RA, Ames BN. Ascorbic acid protects against endogenous oxidative DNA damage in human sperm. *Proc Natl Acad Sci U S A* 1991;88:11003-6.
31. Hughes CM, McKelvey-Martin VJ, Lewis SE. Human sperm DNA integrity assessed by the Comet and ELISA assays. *Mutagenesis* 1999;14:71-5.
32. Beetstra S, Thomas P, Salisbury C, Turner J, Fenech M. Folic acid deficiency increases chromosomal instability, chromosome 21 aneuploidy and sensitivity to radiation-induced micronuclei. *Mutat Res* 2005;578:317-26.
33. Wang X, Thomas P, Xue J, Fenech M. Folate deficiency induces aneuploidy in human lymphocytes in vitro-evidence using cytokinesis-blocked cells and probes specific for chromosomes 17 and 21. *Mutat Res* 2004;551:167-80.
34. Eskes TK. Abnormal folate metabolism in mothers with Down syndrome offspring: review of the literature. *Eur J Obstet Gynecol Reprod Biol* 2006;124:130-3.
35. Courtemanche C, Huang AC, Elson-Schwab I, Kerry N, Ng BY, Ames BN. Folate deficiency and ionizing radiation cause DNA breaks in primary human lymphocytes: a comparison. *FASEB J* 2004;18:209-11.

36. Pogribny IP, Basnakian AG, Miller BJ, Lopatina NG, Poirier LA, James SJ. Breaks in genomic DNA and within the p53 gene are associated with hypomethylation in livers of folate/methyl-deficient rats. *Cancer Res* 1995;55:1894-901.
37. Blount BC, Mack MM, Wehr CM, MacGregor JT, Hiatt RA, Wang G *et al.* Folate deficiency causes uracil misincorporation into human DNA and chromosome breakage: implications for cancer and neuronal damage. *Proc Natl Acad Sci U S A* 1997;94:3290-5.
38. Lopes S, Jurisicova A, Sun JG, Casper RF. Reactive oxygen species: potential cause for DNA fragmentation in human spermatozoa. *Hum Reprod* 1998;13:896-900.
39. Aitken RJ, Baker MA, Sawyer D. Oxidative stress in the male germ line and its role in the aetiology of male infertility and genetic disease. *Reprod Biomed Online* 2003;7:65-70.
40. Kodama H, Yamaguchi R, Fukuda J, Kasai H, Tanaka T. Increased oxidative deoxyribonucleic acid damage in the spermatozoa of infertile male patients. *Fertil Steril* 1997;68:519-24.
41. Wakimoto P, Block G. Dietary intake, dietary patterns, and changes with age: an epidemiological perspective. *J Gerontol A Biol Sci Med Sci* 2001;56 Spec No 2:65-80.
42. Ho E, Ames BN. Low intracellular zinc induces oxidative DNA damage, disrupts p53, NFkappa B, and AP1 DNA binding, and affects DNA repair in a rat glioma cell line. *Proc Natl Acad Sci U S A* 2002;99:16770-5.
43. Oteiza PI, Clegg MS, Zago MP, Keen CL. Zinc deficiency induces oxidative stress and AP-1 activation in 3T3 cells. *Free Radic Biol Med* 2000;28:1091-9.
44. Evenson DP, Emerick RJ, Jost LK, Kayongo-Male H, Stewart SR. Zinc-silicon interactions influencing sperm chromatin integrity and testicular cell development in the rat as measured by flow cytometry. *J Anim Sci* 1993;71:955-62.
45. Yang M, Yang Y, Zhang Z, Hao X, Zheng S, Zhong L *et al.* [Study of zinc in protecting sperm from sodium nitroprusside damage]. *Zhonghua Nan Ke Xue* 2004;10:530-2, 7.
46. Subar AF, Thompson FE, Kipnis V, Midthune D, Hurwitz P, McNutt S *et al.* Comparative validation of the Block, Willett, and National Cancer Institute food frequency questionnaires : the Eating at America's Table Study. *Am J Epidemiol* 2001;154:1089-99.
47. Mares-Perlman JA, Klein BE, Klein R, Ritter LL, Fisher MR, Freudenheim JL. A diet history questionnaire ranks nutrient intakes in middle-aged and older men and women similarly to multiple food records. *J Nutr* 1993;123:489-501.
48. Block G, Thompson FE, Hartman AM, Larkin FA, Guire KE. Comparison of two dietary questionnaires validated against multiple dietary records collected during a 1-year period. *J Am Diet Assoc* 1992;92:686-93.
49. Barratt CL, Aitken RJ, Bjorndahl L, Carrell DT, de Boer P, Kvist U *et al.* Sperm DNA: organization, protection and vulnerability: from basic science to clinical applications--a position report. *Hum Reprod*;25:824-38.

**Figure legend:****Figure 1: Older men with poor micronutrient intake have higher sperm DNA damage**

Sperm DNA damage is reported as alkaline % tail DNA with adjusted means and SE; age and nutrient intake are reported as medians. Low vitamin intake defined as less than the median intake for entire of AGES population. Median value for vitamin C is 162mg, median value for vitamin is 23.7mg for ATE, and median value of zinc is 12.3 mg.

<sup>a</sup>  $p < 0.01$  for old/high vitamin C intake and young/high vitamin C intake vs. referent group (old/low vitamin C intake);  $p < 0.05$  for young/low vitamin C intake vs. referent group.

<sup>b</sup>  $p < 0.10$  for old/high vitamin E intake and young/low vitamin E intake vs. referent group (old/low vitamin E intake);  $p < 0.05$  for young/high vitamin E intake vs. referent group

<sup>c</sup>  $p < 0.05$  for all groups compared to referent group (old/low zinc intake).